Estimation activity and partial purification of Ceruloplasmin from sera of patients with chronic renal Failure and healthy subjects

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Abstract: Ceruloplasmin CP (EC1.16.3.1) activity has been assayed in (32) serum samples of patients with Chronic Renal Failure CRF, and (32) serum samples of healthy individuals without any clinically detectable diseases have been used as control group. The aim of this study is to measure CP activity and partially purifying the enzyme from sera of patients with CRF and healthy individuals. The results of this study revealed that CP activity of patient’s serum shows a highly significant decrease (p < 0.05) compared to that of the healthy individuals. Ceruloplasmin enzyme was partially purified from the serum of patient with CRF and and healthy individual by three steps which operates throughout under mild conditions 4°, acetate buffer (pH 6.8): First step including the use of ammonium sulfate precipitation, second step utilize dialysis with acetate buffer, while the third step consists of gel-filtration on Sephacryl S200(27×1.6cm) . Insufficiently purified ceruloplasmin fractions obtained at various stages are diluted to the appropriate ionic strength and re-utilized. Two proteinous components had been isolated by gel filtration chromatography from the supernatant produced by ammonium sulfate. According to Poly Acryl Amide Electrophoresis PAGE appear two bands by protein staining. Abbreviation: ceruloplasmin, chronic renal failure, ammonium sulfate, dialysis, electrophoresis, gel filtration.

Keywords: Ceruloplasmin, Chronic Renal Failure, Ammonium Sulfate, Dialysis, Electrophoresis, Gel Filtration

1. Introduction

Chronic renal failure (CRF) is a major public health problem with increasing incidence (1). Chronic renal failure characterized by decline in glomerular filtration perturbation of extracellular fluid volume, electrolytes and acid base homeostasis and retention of nitrogenous waste from protein catabolism (2). However, data on the burden of CRF in the Arab world remains poorly understood (3). The primary cause leading to renal failure is a free radical mediated injury to the endothelial cells in the outer medulla. (4). Peroxidation of polyunsaturated fatty acids in the cell membrane generate free radicals and subsequent generation of further unstable radicals leading to a chain of events. This attack makes cell membrane leaky and the functions of absorption and secretion are lost that finally leads to cell death(5). The balance between pro oxidant and antioxidant capacity is shifted towards an increased oxidative stress in CRF patients(6). Chronic renal failure was described as a state of oxidant/antioxidant imbalance (7). There is increasing evidence about the presence of oxidative stress in CRF patients; this seems to be due to multiple factors including an increase in the production of agents from oxidative metabolism (oxygen- derived substances generated by activated leucocytes, transition metal compounds, and other toxins of different molecular weight), and a decrease in antioxidant defenses. Oxidative stress generated in a physiologic or pathologic way and harms the cellular constituents including membrane lipids, proteins, and DNA(8) and exert cytotoxic effects on cellular components(9). Protein molecules may be attacked whenever free radicals are generated (10,11). Ceruloplasmin CP (EC 1.16.3.1) is acute phase reactant protein with a molecular weight of approximately 132 kDa, circulates in plasma as a major copper ion transporter and accounts for more than 95% of the copper found in serum (12). It is one group of serum protein which rises after any form of tissue injury (13). Ceruloplasmin has diverse functions as a copper transporter, it is essential
for iron homeostasis, is involved in tissue angiogenesis and blood coagulation\(^{(14)}\). It has been reported that CP catalyzed the oxidation of Fe (II) to Fe (III), the catalytic cycle involving four of the six copper atoms associated with CP that employ dioxygen as the terminal electron acceptor without the intermediacy of a partially reduced oxygen species such as \(O_2^- \) or \(H_2O_2\)\(^{(15)}\). This oxidase activity increases during inflammation, infection, and injury which suggests that serum CP possibly acts as an antioxidant and as an acute phase protein\(^{(16)}\). Ceruloplasmin may function also as an antioxidant in two different ways: free copper and iron ions are powerful catalysts of free radical damage, by binding copper, CP prevents free copper ions from catalyzing oxidative damage\(^{(17)}\). It has been implied that substantial CP inactivation may occur and free copper ions may be released during exposure to oxidative stress\(^{(18,19)}\). Therefore, damaged CP may cause the augmentation of free radical-mediated damage to other macromolecules upon exposure to oxidative stress\(^{(20)}\). The aim of this study is to measure CP activity and partially purifying the enzyme from sera of patients with renal failure and healthy individuals.

2. Materials and Methods

2.1. Chemicals

All laboratory chemicals and reagents were of analar grade: Tris (Hydroxy methyl) amino methane was obtained from Fluka- Switzerland company, and bovine serum albumin(BSA)from Sigma- USA company.

2.2. Specimens

Thirty two serum samples collected from healthy individuals (20) men and (12) women without any detectable diseases, age range between (20-70) years, and (32) patients serum with chronic renal failure (17) men and from (15) women, age range between (18-75) years. The disease were diagnosed by specialist doctors in AL-emamain hospital, Baghdad, Iraq.

About 10ml of vinous blood was collected in plane tube using plastic disposable syringes and left for 20 minutes at room temperature (25ºC). After coagulation, sera were separated by centrifugation at 1500 xg for 15 minutes. Sera were frozen in -20ºC freezer until analysis.

2.3. Ceruloplasmin Activity Assay

The enzymatic assay of ceruloplasmin oxidase activity was carried out using the modified Rice method\(^{(21)}\) by using p-phenylene diamine-2HCl as a substrate. The CP activity was expressed in term of (U/L).

2.4. Total Serum Protein

Serum protein concentration was determined by Lowry et. al. method\(^{(22)}\), by using bovine serum albumin (BSA) as a standard protein.

2.5. Purification Procedures

Serum was treated with (NH\(_4\))\(_2\)SO\(_4\) as mentioned by Dixon and Webb\(^{(23)}\) in two steps. First, solid (NH\(_4\))\(_2\)SO\(_4\) was added gradually to the serum to 60% saturation in an ice bath, then the mixture was stirred for 15 min, and left at 4 ºC for 60 min. The precipitate was removed by centrifugation. In the second step, the supernatant was treated with solid (NH\(_4\))\(_2\)SO\(_4\) to 80% saturation, stirred for 15 min, and left at 4 ºC for 60 min. The supernatant with ceruloplasmin activity was centrifuged at 2000 xg for 30 min and dialyzed at 4 ºC against the acetate buffer(0.4M, pH= 5.2) overnight. Gel chromatography on Sephadryl S200 was carried out in acetate buffer (0.4M, pH= 5.2) on a column of dimensions (26 x 1.6 cm). The fraction with ceruloplasmin activity was dialyzed against sucrose.

3. Electrophoresis

Polyacrylamide gel electrophoresis (PAGE) was performed as described by Laemmli\(^{(24)}\) using a 12% separating gel, 3 % stacking gel, but without sodium dodecyl sulfate and mercapto ethanol. The protein bands were localized with Coomassie brilliant blue R250.. In order to locate the position of CP bands, polyacrylamide gel was cut to two pieces: the first was stained using staining protein method. They were aligned with the second unstained portion of the gel, which is used to determined CP activity using PPD (P-phenylenediamine) as a substrate\(^{(21)}\).

4. Results

Table 1 reports data of the Ceruloplasmin activity and protein concentration in the sera of CRF patients and healthy individuals compared with the healthy individuals:

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Chronic Renal Failure Patients (n=32) mean±SD</th>
<th>Healthy Individuals (n=32) mean±SD</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ceruloplasmin U/L</td>
<td>74.76±11.3</td>
<td>92.35±20.9</td>
<td>&lt; 0.05</td>
</tr>
<tr>
<td>Protein g/dl</td>
<td>5.66±0.7</td>
<td>7.59±0.41</td>
<td>&lt; 0.05</td>
</tr>
</tbody>
</table>

From table 1 the results above indicated that ceruloplasmin activity and protein concentration were decreased in patients with CRF compared to healthy individuals.

The results of purification experiments are summarized in tables 2 and 3 for CRF patients and healthy individuals, respectively. After each purification step, the protein content and the enzyme activity were determined.
Israa Ghassan Zainal: Estimation Activity and Partial Purification of Ceruloplasmin from Sera of Patients with Chronic Renal Failure and Healthy Subjects

Table (2). Purification of ceruloplasmin from human serum CRF patients.

<table>
<thead>
<tr>
<th></th>
<th>Volume/ml</th>
<th>Total protein mg</th>
<th>Total activity U</th>
<th>Specific activity U/mg</th>
<th>Yield/%</th>
<th>Purification (fold)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crude serum</td>
<td>3ml</td>
<td>129</td>
<td>239.34</td>
<td>0.618</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ammonium sulfate</td>
<td>2ml</td>
<td>61.2</td>
<td>57.9</td>
<td>0.946</td>
<td>72.6</td>
<td>1.5</td>
</tr>
<tr>
<td>Dialysis</td>
<td>1.5ml</td>
<td>30.9</td>
<td>38.03</td>
<td>1.23</td>
<td>47.7</td>
<td>1.2</td>
</tr>
<tr>
<td>Gel filtration</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fraction 1</td>
<td>12ml</td>
<td>3.24</td>
<td>22.2</td>
<td>6.852</td>
<td>27.8</td>
<td>11.1</td>
</tr>
<tr>
<td>Fraction 2</td>
<td>8ml</td>
<td>0.8</td>
<td>22.7</td>
<td>28.38</td>
<td>28.5</td>
<td>45.9</td>
</tr>
</tbody>
</table>

*ammonium sulfate supernatant, \(^{b}\)ammonium sulfate pellet.

Table (3). Purification of ceruloplasmin from human serum healthy subjects.

<table>
<thead>
<tr>
<th></th>
<th>Volume/ml</th>
<th>Total protein mg</th>
<th>Total activity U</th>
<th>Specific activity U/mg</th>
<th>Yield/%</th>
<th>Purification (fold)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crude serum</td>
<td>3ml</td>
<td>174</td>
<td>106.3</td>
<td>0.611</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ammonium sulfate</td>
<td>2ml</td>
<td>96</td>
<td>79.92</td>
<td>0.833</td>
<td>75.2</td>
<td>1.4</td>
</tr>
<tr>
<td>Dialysis</td>
<td>1.5ml</td>
<td>46.05</td>
<td>53.54</td>
<td>1.163</td>
<td>50.4</td>
<td>1.9</td>
</tr>
<tr>
<td>Gel filtration</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fraction 1</td>
<td>12ml</td>
<td>7.8</td>
<td>32.4</td>
<td>4.154</td>
<td>30.5</td>
<td>6.8</td>
</tr>
<tr>
<td>Fraction 2</td>
<td>8ml</td>
<td>3.2</td>
<td>35.7</td>
<td>11.2</td>
<td>33.6</td>
<td>18.3</td>
</tr>
</tbody>
</table>

*ammonium sulfate supernatant, \(^{b}\)ammonium sulfate pellet.

Results of the chromatographic procedure were shown in figures 1&2 for CRF patients and healthy individuals, respectively.

![Figure 1](image1.png)

**Figure (1).** Gel chromatography on a Sephacryl S200 column; protein concentration (blue), ceruloplasmin activity in the sera of human patients (red).

![Figure 2](image2.png)

**Figure (2).** Gel chromatography on a Sephacryl S200 column; protein concentration (blue), ceruloplasmin activity in the sera of human control (red).

According to the present results, ammonium sulphate precipitation to 60-80 % saturation eliminated (27.4% for patient serum and 24.8% for healthy individual serum) of the ballast proteins. From figures 1&2 its clear that there were 2 peaks for enzyme after passing the dialysis sample in the column of gel chromatography on Sephacryl S 200, 72.2% & 69.5% of the ballast proteins for fraction 1 (patient & healthy individual) and 71.5% patients and 66.4% healthy individual of the ballast proteins for fraction 2 were discarded with a loss of ceruloplasmin activity of about 70 % for fractions 1&2 for patients and healthy individuals. Results of this chromatographic procedures were shown in Figures (1 and 2).

The total purification achieved by the procedure outlined in table 1 was (11.1 fold with yield 27.8 % fraction 1 & 45.9 fold with yield 28.5% fraction 2) for patients and (6.8 fold with yield 30.5% fraction 1 & 18.3 fold with yield 33.6% fraction 2) for healthy individuals. The specific activity of the purified enzyme for each fraction increased after using gel filtration steps as clear in tables 1 & 2. The degree of purification of the CP enzyme in every purification step was examined by PAGE as shown in Figure 3.

![Figure 3](image3.png)

**Figure (3).** Protein staining of conventional – PAGE 12% profile of partially purified and crude sera CP. C= control, P=patient, C(A)=control serum using ammonium sulfate, P(A)=patient serum using ammonium sulfate, C(G1)= control serum using dialysis, P(G1)= patient serum using dialysis, C(G1)= control serum using gel filtration fraction1, C(G2)= control serum using gel filtration fraction2, P(G1)= patients serum using gel filtration fraction1 and P(G2)= patients serum using gel filtration fraction2.
5. Discussion

This study is aimed to study the level of CP and attempt to partially purify enzyme in the sera of patients with CRF compare to healthy individuals. Fragmentation of CP during purification and storage has hampered studies of its structure, proteins and majority of the hydrophobic protein precipitate produced by polyethylene glycol (4000). It was achieved by the procedure outlined in tables (6). Serum using polyethylene glycol (4000). After gel chromatography on Sephacryl-S200 column, two fractions were appeared in patient (27.8 & 28.5)% & in healthy individual (30.5&33.6)% of the ballast proteins were discarded. Results of this chromatographic procedure were shown in figures (1&2) and tables (1&2). The total purification achieved by the procedure outlined in tables (1&2) were (11.1 & 45.9) fold for patient and (6.8&18.3) fold for healthy individual over the first soluble fraction. The specific activity of the purified enzyme was (6.852&28.38) U/mg for patients fragments and (4.154&11.2) U/mg for healthy individual fractions, in literature specific activity were (6.8&13.9) U/min/mg for healthy individual only (41). Ahmed and AL-Helaly (41) had been isolated two proteinous components by gel filtration chromatography from the precipitate produced by polyethylene glycol (4000). It was found that only the second peak had a high activity for CP.

References

94 Israa Ghassan Zainal: Estimation Activity and Partial Purification of Ceruloplasmin from Sera of Patients with Chronic Renal Failure and Healthy Subjects


