Antioxidant Activity of Bioactive Protein of Kerang Kepah (*Atactodea striata*) from South Sulawesi

Tahirah Hasan¹, Abdul Wahid Wahab¹, Natsir Djide², Muhammad Zakir^{1,*}

¹Department of Chemistry, Graduate School of Science, Hasanuddin University, Makassar 90245, South Sulawesi, Indonesia

²Faculty of Pharmacy, Hasanuddin University, Makassar 90245, South Sulawesi, Indonesia

Email address:

tahirahhasan@yahoo.com (T. Hasan), wahidwahab@yahoo.com (A. W. Wahab), natsirjide@yahoo.com (N. Djide), muhammadzakir@gmail.com (M. Zakir)

To cite this article:

Tahirah Hasan, Abdul Wahid Wahab, Natsir Djide, Muhammad Zakir. Antioxidant Activity of Bioactive Protein of Kerang Kepah (*Atactodea striata*) from South Sulawesi. *American Journal of Biomedical and Life Sciences*. Vol. 3, No. 6, 2015, pp. 111-114.

doi: 10.11648/j.ajbls.20150306.12

Abstract: Antioxidant activity test on protein extract of Kerang Kepah (*Atactodea striata*) by scavenging method of free radical of DPPH (N, N-diphenyl-N-picrylhydrazyl) has been carried out. This study was conducted to determine the antioxidant activity and the amino acid composition of the protein extract of *Atactodea striata*. The analysis results showed that the protein extract of *Atactodea striata* has the most complete amino acid composition and the highest antioxidant activity of the protein fraction at a rate of 30-50% ammonium sulfate saturation with IC₅₀ value of 183.75 µg/mL. However, when it is compared to the antioxidant activity of ascorbic acid (IC₅₀ values = 2.51 µg/mL), the antioxidant activity of the protein extract fraction of *Atactodea striata* is still much lower.

Keywords: *Atactodea striata*, Antioxidant, DPPH, HPLC

1. Introduction

Consumption of sea shells as a popular sea food in the last few decades is increasing. High diversity of marine organisms, including sea shells, is a pillar of traditional knowledge of the various drugs. Mollusks a marine commodity which is a potential source of bioactive compounds as candidates for various purposes. Bivalve and gastropod mollusc groups whose existence is quite abundant in the tropical waters as a source of animal protein has a relatively cheap prices. Bioactive compounds found in mollusks which are identified as peptide, dipeptides, sesquiterpenes, squalene, terpenes, alkaloids, polipropionate, nitrogen compounds, fatty acid derivatives, makrolide and other compounds have a specific bioactivity [1, 2, 3].

Several studies have been conducted on the bioactive compounds of molluscs, especially bivalves and gastropods that is potential as a nutraceutical or pharmaceutical. Some of them are marine leeches *Discodoris* sp. [5], snails *Ipongi* *Salmo fascilaria* [6], *Cyclina sinensis* [7], abalone *Haliotis discus hannai* Ino [8], and green mussels *Perna viridis* Linn [9].

Natural products isolated from bivalves and gastropods

have been used, among others, as an antioxidant, antitumor, antiviral, antibacterial, antifungal, anticancer, cytotoxic and enzyme inhibitor [8, 9]. However, it is still lack a report so far on the antioxidant activity of the protein metabolites isolated from the class of bivalves.

This study was conducted to isolate the bioactive proteins of Kerang Kepah (*Atactodea striata*) from waters of Laiya Island, Pangkep, South Sulawesi. Active compounds obtained will be tested on their activity as an antioxidant. The results of this study will be developed and applied in the biotechnology industry for further utilization in the formal health care system. This study is also important in the exploration of active compounds as antioxidants in other marine resources.

One of marine organisms that attracts attention is Kerang Kepah (*Atactodea striata*). This type of shell fish is a food that favored people since long time ago, and even the shell is used as traditional medicine for a variety of chronic diseases, especially for hepatitis. Therefore, *Atactodea striata* is suspected to contain bioactive compounds (primary and secondary metabolites) because traditionally it has been used as medicine.

Several researches indicate that some species of mollusc has a protein content so that they continue to grow and

* Muhammad Zakir: Corresponding author

develop properly at sea where a lot of bacteria and other predators around them. The protein content of marine organism can act as an agent for reducing free radicals, chelating metal ions, trapping free radicals, and can inhibit some types of bacteria. However, there is still less research data on protein compounds as raw material medicines. Meanwhile, the protein has several advantages as a very promising material for antimicrobial drugs and anticancer acceptable without any side effects; so that researchers began to develop a treatment or therapy of disease by using a protein compound [10].

Based on the above cases, this research is conducted to assess the active protein compound of marine shells from Laiya Island Pangkep, South Sulawesi. The purpose of this research is to determine the antioxidant activity and the amino acid composition of protein extracts of Kepah (*Atactodea striata*). The result of this research is expected to reveal the potential of sea shells as a producer of compounds that have antioxidant activity.

2. Methods

2.1. Sample Preparation

Samples were taken in coastal waters of Laiya Island Pangkep, South Sulawesi. Samples were cleaned from dirt, released from the shell, and weighed 500 g and then milled using a blender by adding Tris-HCl buffer 0.1 M pH 8.3 and then stored in a 4°C refrigerator overnight, and finally filtered with a Buchner funnel. The filtrate obtained is freeze-dried and then melted 2-3 times, centrifuged at 6000 rpm at 4°C for 30 min, and finally the supernatant is stored in the refrigerator before continuing trials of antioxidant activity.

2.2. Fractionation with Ammonium Sulfate

Crude protein extract is further isolated by ammonium sulfate fractionation. The saturation level (0-90%) based on the method of Bollag and Edelstein [11]. The resulting protein precipitate is dissolved in a solution of Tris-HCl buffer pH 8.3, and followed by dialysis using selophan bag (Sigma).

2.3. Dialysis

Dialysis was performed using a cellophane membrane (Sigma D 0655). According to Plummer [12], deposition and fractionation results of each level of saturation of ammonium sulfate is dialyzed in a 0.1 M Tris-HCl pH 8.3, then dialyzed with Tris-HCl buffer pH 8.3 0.01M. Dialysis continued until the buffer solution is colorless. Protein content of dialysate was tested as well as antibacterial and antioxidant activity.

2.4. Determination of Protein and Amino Acid Composition

Determination of protein levels were calculated using a Bio-rad. In Eppendorf tube, it is inserted 4 µL of sample solution and added 746 µL and 200 µL dH₂O. Bio-rad protein assay was homogenized and allowed to stand for 30 minutes. Absorbance is measured at the maximum wavelength.

Determination of protein concentration is carried out using linear regression equation and the amino acid composition through HPLC method.

2.5. Antioxidant Activity Test Method Immersion Effect Against Free Radical DPPH

Antioxidant activity test was conducted by soaking effect on DPPH free radicals. Antioxidant compounds tested its effectiveness in reducing the DPPH free radical activity. Protein extracts made some concentration (ppm), respectively included in 5 mL volumetric flask, add 1.0 mL into each 0.1 mM DPPH solution and the volume was made up to the mark, then incubated at 37 °C for 30 min and the absorbance was measured at a wavelength of 517 nm. Ascorbic acid is used as a comparison.

3. Results and Discussions

Kepah Samples (*Atactodea striata*) obtained from coastal waters of Laiya Island, Pangkep Regency, South-Sulawesi. Meat samples of *Atactodea striata* was extracted using Tris-HCl buffer 0.1 M pH 8.3 and then stored in a 4°C refrigerator overnight and filtered with a Buchner funnel. The filtrate obtained was frozen and melted 2-3 times and then centrifuged at 6000 rpm at 4°C for 30 minutes. Crude extract in protein isolation results was further fractionated with ammonium sulfate with the saturation level (0-90%) based on the method of Bollag and Edelstein [11]. The resulting protein precipitate was dissolved in a solution of Tris-HCl buffer pH 8.3, followed by dialysis using selophan bag (Sigma D 0655).

According to Plummer [12], precipitation of fractionation results of each level of ammonium sulfate saturation is dialyzed in a 0.1 M Tris-HCl pH 8.3, and further dialyzed with Tris-HCl buffer pH 8.3 0.01M. Dialysis is continued until the buffer solution is colorless. Dialysate obtained was tested for protein content and antioxidant activity.

Determination of protein levels were calculated using a Bio-rad. Eppendorf tube inserted into a 4 µL sample solution was added 746 µL and 200 µL dH₂O. Bio-rad protein assay was homogenized and allowed to stand for 30 minutes. Absorbance is measured at the maximum wavelength. Determination of protein content is carried out using a linear regression equation. Measurement results of protein content of Kepah (*Atactodea striata*) using Bio-rad can be seen in Table 1.

Table 1. The protein concentration of the fractionation of various levels of ammonium sulfate saturation of Kepah extract (*Atactodea striata*).

No	Protein fraction	Fraction volume (mL)	Protein concentration (mg/mL)	Total protein (mg)
1	Crude extract	500	2.015	1.007
2	F ₁ (30-50 %)	420	1.48	621.6
3	F ₂ (30-50 %)	400	1.320	528
4	F ₃ (50-70 %)	370	0.800	296
5	F ₄ (70-90 %)	200	0.640	128

The results of the analysis of protein content of Kepah

(*Atactodea striata*) in Table 1 indicate that the protein concentration of the crude extract is 1,007 mg. Distribution of protein in each fractionation with ammonium sulfate at a rate of 0-90% saturation showed the highest protein concentration was found in the fraction with 30% saturation level is 621.6 mg. While the lowest protein concentration found in the fraction with 90% saturation level is 128 mg. The measurement results showed that the total protein content of each fraction proteins differ from each fraction. This suggests that the protein precipitated from each fraction is a different protein. The protein is precipitated by differences in solubility in water [13].

The results of the analysis of the amino acid composition of protein extract fraction of Kepah (*Atactodea striata*) by HPLC (High Performance Liquid Chromatography) can be seen in Table 2.

Table 2. Amino Acid Composition of Protein Extract Fraction Kepah (*Atactodea striata*) with HPLC Method.

Amino acid	Protein extract			
	F ₁ (mg/100 g)	F ₂ (mg/100 g)	F ₃ (mg/100 g)	F ₄ (mg/100 g)
L-Histidin	-	-	-	-
L-Serin	31.26	40.79	-	34.75
L-Arginin	37.21	55.64	-	-
Glysin	60.59	172.20	275.79	31.89
L-Asam Aspartat	47.53	24.08	-	-
L-Asam Glutamat	178.58	106.67	-	124.78
L-Treonin	23.76	65.47	-	-
L-Alanin	37.73	97.38	-	-
L-Prolin	28.30	34.19	-	-
L-Sistin	-	30.99	-	-
L-Lisin	30.15	505.59	-	-
L-Tirosin	49.02	208.90	-	-
L-Metionin	484.23	245.61	477.93	350.04
L-Valin	49.75	42.16	-	-
L-Isoleusin	19.36	54.01	-	-
L-Leusin	38.08	39.92	-	-
L-Phenilalanin	11.60	65.15	-	72.72

Antioxidant activity test was conducted using the damping effect against free radical DPPH using visible spectrophotometer at a wavelength of 517 nm. Protein extracts was tested for its effectiveness in reducing the DPPH free radical activity. Ascorbic acid is used as a comparison. Reduction of DPPH free radical activity of each protein fraction, with a respective ammonium sulfate saturation level (%) 0-30, 30-50, 50-70 and 70-90, can be seen in Table 3.

It is clearly seen from the antioxidant activity of each protein fraction that the percent of DPPH with the greatest antioxidant activity is found in fraction F₂ with IC₅₀ 183.75µg/mL, but it is still far below the antioxidant activity of ascorbic acid which has IC₅₀ 2.51µg/mL. The significant difference in the antioxidant activity of each fraction is probably resulted from differences in the amino acid composition of each fraction [14, 15].

Table 3. Antioxidant activity of Protein Extract Fraction Kepah (*Atactodea striata*) with method Damping Effects Against Free Radical DPPH.

Protein fraction	M (µg/mL)	% Inhibition	IC ₅₀ (µg/mL)
F ₁ (0-30 %)	50	26.71	383.71
	100	30.16	
	150	33.60	
	200	37.30	
	250	40.74	
F ₂ (30-50 %)	50	43.65	183.75
	100	46.56	
	150	47.08	
	200	51.59	
	250	53.17	
F ₃ (50-70 %)	50	16.40	906.05
	100	19.31	
	150	20.11	
	200	22.49	
	250	24.34	
F ₄ (70-90 %)	50	18.62	688.33
	100	21.96	
	150	25.13	
	200	27.25	
	250	28.04	
Ascorbic acid	5	54.92	2.51
	10	61.66	
	15	78.24	
	20	83.94	
	25	92.22	

4. Conclusions

Based on the results of research conducted, it can be concluded that the protein extract of Kepah (*Atactodea striata*) at the level of 30-50% saturation of ammonium sulfate has the highest antioxidant activity against DPPH (diphenyl picrylhydrazyl) free radical reduction with IC₅₀ value of 183.75µg/mL. This analysis result with DPPH free radical reduction activity showed that the protein fraction of *Atactodea striata* is considered as an effective antioxidant.

References

- [1] J.L. Balcazar, R.Z. Blas, D. Cuningham, Vendrell and Muzquiz, The Rute of Probiotic in Aquaculture, Veterinary Microbiol. vol. 114, pp. 173-186, 2006.
- [2] J.W. Blunt, B.R. Copp, M.H.G. Munro, Northcote and M.R. Prinsep, Marine Natural Products. Natural Product Reports, vol. 23, pp. 26-78, 2006.
- [3] U. Grienke, J. Silke, D. Tasdemis, "Bioactive compounds from marine mussels and their effects on human health," Food Chem. Vol. 142, pp. 48-60, 2014.
- [4] R.D. Barnes, Invertebrate Zoology, Saunders College, Fourth Edition, 1982.
- [5] L. Nurjanah, D. Hardjito, Monintya, M. Bintang, and D.R. Agungpriyono, "Aktivitas antioksidan lintah laut (*Discodoriss*) dari perairan Pulau lintah laut dari perairan Pulau Buton Sulawesi Tenggara," Prosiding Seminar Nasional Pengolahan Produk dan Bioteknologi Kelautan dan Perikanan. Jakarta, 2009.

- [6] A. Nurjanah, Abdulla, and A. Apriand, "Aktivitas antioksidan dan komponen bioaktif pada keong ipong-ipong (*Fasciolaria salmo*),"J. Pengolahan Hasil Perikanan Indonesia, vol.14, pp 22-29, 2011.
- [7] C. Jiang, M. Wang, J. Lin, D. Gan, X. Zeng, "Extraction, preliminary characterization, antioxidant and anticancer activities in vitro of polysaccharides from *Cyclinosinensis*," Carbohydrate Polymers, vol 84, pp. 851-857, 2011.
- [8] D.Y. Zhou, B.W. Zhu, L. Qiao, H.T. Wu, D.M. Li, J.F. Yang and Y. Murata, "In vitro antioxidant activity of enzymatic hydrolysates prepared from abalone (*Haliotis discus hannai*no) viscera," Food and Bioproducts Processing, 2011.
- [9] V.N. Madhu, K. Sivaperumal, K. Kamala and B.G. Kulkarni, "Antibacterial and Antioxidant Activities of The Tissue Extract of *Verna viridis* From Versova Coast," International Journal of Pharmacy and Pharmaceutical Sciences, Mumbai, 2014.
- [10] L. Huang, Protein dalam Air Mata Obat untuk AIDS, (On line), (<http://www.rad.net.id/warta/wao4701.htm>, (diakses 2 Januari 2013), 1999.
- [11] Nollet, M. L. Leo, "Handbook of Food Analysis and Amino Acid," 1996.
- [12] D.T. Plummer, "An Introduction to Practical Biochemistry," Second Edition. Tata Mc. Graw Hill Publishing Company Ltd. New Delhi, 1979.
- [13] R.K. Scopes, "Protein Purification Principles and Pactice," Second edition, Springer-Verlag, New York, 1987.
- [14] Liyana, F. Pathiranam, Shahidi," Antioxidant activity of commercial soft and hard wheat (*Triticum aestivum* L.) as affected by gastric pH conditions," J. Agric. Food Chem. Vol. 53, pp. 2430-2440. 2005.
- [15] W. Moka,"Identifikasi dan Inventarisasi Jenis Kerang Laut yang Digunakan Sebagai Obat Tradisional di Sulawesi Selatan", UNHAS, Makassar, 1982.