Effect of Huoxue Jiedu Jiangtang Formulation on Endoplasmic Reticulum Stress in Diabetic Atherosclerosis

Fu Xianzhao*, Huang Wenhua, Li Chunyan, Li Xingchan, Qiu Haixian, Liang Liudan, Cao Qiuxia, Liang Liuyue

Clinic Medical College of Youjiang Medical National College, Baise, China

Email address: 1620100638@qq.com (Fu Xianzhao)
*Corresponding author


Received: October 31, 2018; Accepted: December 7, 2018; Published: December 27, 2018

Abstract: Objective: To study the effects of Huoxue Jiedu Jiangtang formulation (HJJF) on endoplasmic reticulum stress (ERS) coupled inflammatory response. From the molecular mechanism in diabetic vascular disease. Methods: Type 2 diabetes were established in male SD rats by feeding high lipid diet and injection of streptozotocin. After 2DM models established successfully, the rats were divided into model group, low-dose HJJF group (HJJF1), high-dose HJJF group (HJJF2), and Western medicine group (Gliquidone+Benazepril), and accepted corresponding drugs for 8 weeks respectively. Another normal group were used as control group, fed with normal diet, no drug intervention. The levels of fasting blood glucose (FBG), serum superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), IL-6 and TNF-α were tested. The ERS signaling molecules glucose-regulated protein 78 (GRP78) and c-Jun N-terminal kinases (JNK) mRNA transcription level in thoracic aorta vessel were determined by reverse-transcription polymerase chain reaction (RT-PCR). Results: Compared with model group, After drugs intervention, all administered groups could significantly decrease FPG (P<0.05), elevate SOD and GSH-Px (P<0.05); all could reduce levels of TC, TG, LDL-C, IL-6 and TNF-α (P<0.05), and raise HDL-C (P<0.05); all could depress the transcription levels of GRP78 and JNK mRNA (P<0.05). With increasing dose, the HJJF effect is more significant (P<0.05). Conclusion: HJJF could improve insulin resistance, correct lipid metabolism disorders, and enhance the antioxidative ability, inhibit the response of ERS coupled inflammatory, showing the multi-target treatment characteristics of Traditional Chinese Medicine.

Keywords: Diabetes, Atherosclerosis, Huoxue Jiedu Jiangtang Formulation, Inflammatory Reaction, Endoplasmic Reticulum Stress

1. Introduction

Diabetic atherosclerosis (AS) is a chronic inflammatory-related metabolic disease, and Endoplasmic reticulum stress (ERS) is a bridge between metabolic abnormalities and inflammatory reactions. In 2004, Ozcan U et al. [1] published an article in «Science», pointing out that ERS is a common pathway of multiple stressors and an important link in the occurrence and development of insulin resistance and complications in diabetes mellitus. In 2016 «Nature», Wang Met et al. [2] explained that ERS contributes to the aetiology of many human diseases. Therefore, ERS is expected to become a new target for the treatment of diabetes mellitus. Although the pathogenesis of diabetic AS is complex, the pathological state of diabetes mellitus, such as high glucose, high fat, oxidative stress and inflammatory reaction, are related to ERS, which link the metabolic abnormalities and the development of diabetes mellitus, can cause apoptosis of macrophages and smooth muscle cells and participate in the occurrence and development of diabetic AS [3]. According to traditional Chinese medicine (TCM) theory, diabetes and coronary heart disease belong to the category of Xiao Ke and chest obstruction respectively, whose pathogenesis are deficiency of Qi and Yin, dry and heat holding in store, and with the passing of time, the fluid was burnt into phlegm, Yin deficiency leads to Yang insufficiency, blood coagulate into stasis, and finally, the accumulated phlegm and stasis turn to...
toxin. Therefore, deficiency of Qi and Yin, accumulation of blood stasis toxin are the main pathogenesis of diabetes and chest obstruction [4]. Previous clinical studies have found that Huoxue Jiedu Jiangtang formulation (HJJF), with the functions of Nourishing yin and replenishing qi, activating blood circulation and detoxicating, can significantly reduce pro-inflammatory factors, regulate anti-inflammatory/pro-inflammatory balance, effectively inhibit inflammation in non revascularization of diabetic acute coronary syndrome (critical coronary heart disease) patients, and improve their clinical curative effect [5]. In this study, animal models were used to investigate the molecular mechanism of ERS coupled with inflammatory response in the development of diabetic AS.

2. Methods

2.1. Model Establishment

120 male SD rats Weighing 220-250 g were fed (each cage hold four rats) at room temperature of 22-25°C and relative humidity of 40-50%. After adapted feeding for 7days, and fasted for 10 hours, the rats were intraperitoneally injected streptozotocin (STZ) (Beijing Hua Yang Biotechnology Co., Ltd.) once 50mg / kg. 72 hours later, blood glucose were tested with blood glucose test strips (Wuhan Boster Biological Technology, LTD). Animals with FBG≥16.7mmol/L were used to investigate the molecular mechanism of ERS coupled with inflammatory response in the development of diabetic AS.

2.2. Grouping and Administration of Rats

Total of 102 rats were successfully modeled, which subsequently were randomly allocated into model group 25, HJJF low dose group (HJJF1) 26, HJJF high dose group (HJJF2) 25, western medicine (Gliquidone+benazepril) group 26. In addition, 25 homology rats were taken as normal group, accepted intraperitoneal injection of normal saline, and fed with ordinary feed. HJJF (Ginseng, Astragalus membranaceus, Ophiopogon japonicus, Cornus officinalis, Rehmannia glutinosa, Rhubarb, turtle shell, peach kernel, dan pi, Coptis chinensis, Salvia miltiorrhiza, yam, Schisandra chinensis, lard 10%, cholesterol 1.5%, porcine bile salt 0.3%, custard powder 10%) provided by Jiangsu synergetic Biolog al Technology, LTD). Animals with FBG≥16.7mmol/L were ceded equal volume solution. HJJF (Gliquidone (Beijing Wan Hui Shuanghe Pharmaceutical Co., Ltd) 7.50mg/kg, which were confected equal volume solution. The specific experimental procedures were carried out according to the instructions of Real-time PCR kit.

2.4.1. Detection of Serum Lipids, SOD, GSH-Px, IL-6 and TNF-a

Total cholesterol (TC), triglyceride (TG), low density lipoprotein cholesterol (LDL-C) and high density lipoprotein cholesterol (HDL-C) were determined by automatic biochemical instrument. SOD content was tested by xanthine oxidase method, interleukin-6 (IL-6), tumor necrosis factor-alpha (TNF-a) and GSH-Px levels were checked by enzyme-linked immunosorbent assay (ELISA). The kits were purchased from Wuhan Doctor De Bioengineering Co., Ltd. The specific experimental procedures were carried out according to the instructions.

2.4.2. Using Real-Time PCR Method to Detect the Transcription of GRP78 and JNK mRNA in Aorta

Taking out the aortas from -80°C refrigerator, grinding in a mortar, extracting the total RNA, conducting electrophoresis in agarose gel, and checking its integrity, turn it into cDNA (at 42°C 50 min, then 95°C 5 min) by reverse transcription. The obtained cDNA was used as template to amplify the target gene by PCR (according to the instructions of Real-time PCR kit). The reference and target gene primers were designed and synthesized by Shanghai Genechem Co., LTD. The sequence of primers are as follows: GAPDH upstream 5’-TTCAACGGCACAGTCA AGG-3’, downstream 5’-CTCAGCACCAGCATCACC-3’, amplified product 114 bp; GRP78 upstream 5’-TTTACGCTGGGCGCTTAC-3’, downstream 5’-TTTACGCTGGGCGCTTAC-3’, amplified product 117 bp; c-JNK upstream 5’-GGATTTGAGGAG CGAACAT-3’, downstream 5’-ACTCTGCTGTCGCTACCG AGGC-3’, amplified product 163 bp. The process of amplification of obtained cDNA is as follows: 96°C 4min, one cycle, subsequently 94°C 30 second, 58°C 30 second, 72°C 30 second, 40 cycle. The number of cycles when the set threshold reached (CT) were recorded, and the specificity of primers was detected by dissolution curve. With 2^△△CT representing the expression of target gene in experimental group, △△CT= Experience group (Ct_target gene−Ct_GAPDH) -control group (Ct_target gene−Ct_GAPDH).
2.5. Statistical Analysis

All data were expressed by mean ± standard deviation (X±S) and SPSS 19.0 statistical software was used to process the data. One-way ANOVA was used to analyze the differences between groups. Homogeneity of variance test was put into application to test the homogeneity of variance. If the variances were homogeneous, LSD was applied to calculate the variances; if not, Tamhane’s T2 is put to use. P<0.05 was considered to indicate the statistically significant difference.

3. Result

3.1. Changes of Experimental Process in Rats

After successful modeled, rats urinated and drank more, with weight loss and withered hair, acting slow and sluggish. But the intragastric administration was successful and no rat died.

3.2. Effects of HJJF on Activities of FBG, SOD, GSH-Px, TNF-α and IL-6 in Rats

The results showed that compared with the model group, in drug treated groups, FBG decreased significantly (P<0.05), antioxidant enzymes SOD and GSH-Px increased (P<0.05), inflammatory factors TNF-α and IL-6 declined (P<0.05); HJJF group had better curative effect than western medicine group, and the effect was more significant with the increasing HJJF dose (P<0.05), but there was no significant difference between HJJF groups (P>0.05). The results are shown in Table 1.

Table 1. Effects of HJJF on FBG, SOD, GSH-Px, TNF-α and IL-6 in rats (X±S).

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>FBG (mmol/L)</th>
<th>SOD (u/mL)</th>
<th>GSH-Px (u/mL)</th>
<th>TNF-α (ng/L)</th>
<th>IL-6 (ng/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>normal group</td>
<td>25</td>
<td>5.19±0.71</td>
<td>391.7±63.2</td>
<td>372.2±489.4</td>
<td>51.64±2.86</td>
<td>301.21±78.87</td>
</tr>
<tr>
<td>model group</td>
<td>25</td>
<td>16.71±1.63</td>
<td>265.2±49.7</td>
<td>430.1±639.7</td>
<td>417.68±63.72</td>
<td>3151.58±857.21</td>
</tr>
<tr>
<td>HJJF1 group</td>
<td>26</td>
<td>10.21±1.34</td>
<td>328.0±51.9</td>
<td>4548.3±489.6</td>
<td>138.75±51.12</td>
<td>912.32±231.32</td>
</tr>
<tr>
<td>HJJF2 group</td>
<td>25</td>
<td>9.77±1.55</td>
<td>358.6±81.7</td>
<td>4811.2±588.6</td>
<td>116.51±45.56</td>
<td>865.45±126.21</td>
</tr>
<tr>
<td>western medicine</td>
<td>26</td>
<td>10.79±1.48</td>
<td>305.1±41.7</td>
<td>4440.3±478.7</td>
<td>141.54±50.27</td>
<td>956.38±261.36</td>
</tr>
</tbody>
</table>

Notes: Compared with the normal group, P<0.01; Compared with the model group, P<0.05; Compared with the western medicine group, P<0.05

3.3. Effect of HJJF on Blood Lipid in Rats

Compared with the normal control group, the contents of TC, TG and LDL-C in the model group increased significantly (P<0.01), and HDL-C decreased significantly (P<0.01); compared with the model group, after administration, the contents of TC, TG and LDL-C decreased, and the contents of HDL-C increased in all drug treated groups (P<0.05). The effect of HJJF was more significant with the increase of dosage, and the difference was statistically significant compared with the model group. There was no significant difference between HJJF groups (P>0.05). The results are shown in Table 2.

Table 2. Effects of HJJF on TC, TG, LDL-C and HDL-C (X±S).

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>TC (mmol/L)</th>
<th>TG (mmol/L)</th>
<th>HDL-C (mmol/L)</th>
<th>LDL-C (mmol/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>normal group</td>
<td>25</td>
<td>1.82±0.11</td>
<td>0.91±0.12</td>
<td>0.93±0.07</td>
<td>0.91±0.52</td>
</tr>
<tr>
<td>model group</td>
<td>25</td>
<td>5.38±0.41</td>
<td>2.31±0.24</td>
<td>0.52±0.06</td>
<td>2.11±0.75</td>
</tr>
<tr>
<td>HJJF1 group</td>
<td>26</td>
<td>2.92±0.31</td>
<td>1.81±0.31</td>
<td>0.78±0.07</td>
<td>1.73±0.23</td>
</tr>
<tr>
<td>HJJF2 group</td>
<td>25</td>
<td>2.81±0.30</td>
<td>1.71±0.22</td>
<td>0.81±0.09</td>
<td>1.61±0.21</td>
</tr>
<tr>
<td>western medicine</td>
<td>26</td>
<td>2.96±0.33</td>
<td>1.88±0.21</td>
<td>0.75±0.08</td>
<td>1.77±0.15</td>
</tr>
</tbody>
</table>

Notes: Compared with the normal group, P<0.01; Compared with the model group, P<0.05; Compared with the western medicine group, P<0.05

3.4. Effects of HJJF on GRP78 and JNK mRNA Transcription Levels in Rat Aorta

According to the result of Table 3, compared with the normal control group, the expression of GRP78 and JNK in the model groups were significantly higher (P<0.01); after administration, the transcription of GRP78 and JNK in each group was lower than that in the model group (P<0.05); compared with the western medicine group, the effect of HJJF2 group was more significant (P<0.05); the effect of HJJF was better with the increase of dosage, but among the HJJF groups. The difference was not statistically significant (P>0.05). The results are shown in Table 3, Figure 1.

Table 3. Relative transcriptional level of GRP78 and JNK mRNA in rats aorta in each group (2-△△CT, X±S).

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>GRP78</th>
<th>JNK</th>
</tr>
</thead>
<tbody>
<tr>
<td>normal group</td>
<td>25</td>
<td>1.05±0.21</td>
<td>1.07±0.26</td>
</tr>
<tr>
<td>model group</td>
<td>25</td>
<td>7.12±0.38</td>
<td>7.63±0.24</td>
</tr>
<tr>
<td>HJJF1 group</td>
<td>26</td>
<td>4.21±0.87</td>
<td>4.82±0.91</td>
</tr>
<tr>
<td>HJJF2 group</td>
<td>25</td>
<td>3.15±0.37</td>
<td>3.22±0.32</td>
</tr>
<tr>
<td>western medicine</td>
<td>26</td>
<td>4.36±0.41</td>
<td>4.94±0.41</td>
</tr>
</tbody>
</table>

Notes: Compared with the normal group, P<0.01; Compared with the model group, P<0.05; Compared with the western medicine group, P<0.05
Figure 1. Amplification and dissolution curves of GRP78 and JNK mRNA in aorta.
4. Discussion

Endoplasmic reticulum (ER) is an organelle for protein and steroid synthesis. When ER homeostasis imbalance leads to the accumulation of unfolded protein and disordered lipid synthesis, it will trigger a series of intracellular stress responses, called ERS [6]. Glucose regulated protein 78 (GRP78) is the feeling molecular for ER homeostasis. When ER is in steady state, GRP78 combined with the domain of three transmembrane proteins: inositol demand enzyme 1 (IRE-1), activating transcription factor 6 (ATF6) and protein kinase R like endoplasmic reticulum kinase (PERK), which exposed to the ER cavity. When severe ERS sustained, GRP78 will dissociate with PERK, ATF6 and IRE-1 as a result of its higher affinity with unfolded proteins, and then, the dissociated PERK, ATF6 and IRE-1 activate downstream cJun ammonia acid terminal kinase (c-JNK), CCAAT enhancer binding protein homologous protein (CHOP), cysteine aspartic proteinase -12 (caspase12) apoptotic signaling pathways, leading to cell apoptosis [7]. In JNK pathway, activated IRE-1 combines with tumor necrosis factor receptor associated factor 2 (TRAF2) through its intracytoplasmic binding domain to form IRE-1-TRAF2 complex, and then activates downstream apoptosis signal regulating kinase 1 (ASK1), and forms IRE-1-TRAF2-ASK1 complex with ASK1, which makes JNK phosphorylated and activated, subsequently, activated JNK inhibits the anti-apoptotic activity of Bcl-2 and enhances the apoptotic function of BIM, also participate in the regulation of caspase-12, phosphorylate CHOP, and increase the apoptotic effect of each other by regulating CHOP activity [7-8]. Activated JNK can also activate activator protein-1 (AP-1) and increase the expression of pro-inflammatory factor genes. IRE1α-TRAF2 complex can also phosphorylate and degrade IkB, release NF-kB into nucleus, activate the transcription of pro-inflammatory factor genes [9-10], lead to the increase of pro-inflammatory factors (TNF-a, IL-6, etc.) and increase the vulnerability of atherosclerotic plaques [11]. A large expression of endoplasmic reticulum chaperone was found in the samples obtained by percutaneous transluminal plaque rotated resection, and ERS apoptotic pathway was activated in unstable plaques [12].

Diabetes mellitus results in impairment of normal pathways of glucose metabolism, overactivation of four bypass pathways, including polyol pathway, protein kinase C (PKC) pathway, advanced glycation end products (AGEs) pathway and hexosamine pathway, which enhances the activity of NADPH oxidase and produces excessive reactive oxygen species (ROS) clusters in cells [13]. The accumulation of ROS affects the homeostasis of endoplasmic reticulum, which hinders the folding of protein spatial structure and induces the accumulation of unfolded proteins to induce ERS [14]. In addition, insulin resistance impairs the function of lipoprotein lipase, decreases the clearance rate of VLDL, and hyperinsulinemia promotes the synthesis of VLDL in the liver, presenting increased TG and LDL, decreased HDL and disordered lipid metabolism. Enhanced oxidative stress, which oxidizes LDL to ox-LDL, can not be recognized by LDL receptors for normal degradation, and has a high affinity with scavenger receptors of macrophages. This process has no negative feedback mechanism, leading to a large accumulation of cholesterol esters in macrophages, triggering ERS, which further exacerbating inflammatory response and leading to a vicious cycle [15]. Therefore, endoplasmic reticulum stress-coupled inflammation plays an important role in the occurrence and development of diabetic atherosclerosis. It has been found that insulin resistance can make macrophages more sensitive to endoplasmic reticulum stress-mediated apoptosis, induce inflammation, macrophage and smooth muscle cell apoptosis, and promote atherosclerosis occurrence and development [16-17].

Antioxidant enzyme system is an important barrier for the body's antioxidant injury, which mainly includes GSH-Px, SOD. GSH-Px is an important peroxidase widely existing in organisms, which is a detoxification enzyme, with the ability to catalyze the conversion of reduced glutathione (GSH) to oxidized glutathione (GSSG), reduce toxic peroxides to non-toxic hydroxyl compounds, promote H2O2 decomposition, remove hydrogen peroxide and other organic peroxides in organisms. SOD is a natural antioxidant enzyme of superoxide anion playing an important role in antioxidant damage. The synergistic effect of GSH-Px and SOD can prevent diabetes from producing excessive ROS.

According to TCM theory, deficiency of both Qi and Yin, dryness-heat accumulation are the basic pathogenesis of diabetic AS. Deficiency of Qi leads to blood stasis, Yin deficiency aggravates internal heat, heat and blood stasis can transform into toxin, phlegm and blood stasis can pile up over time (atherosclerotic plaque) [18]. At the same time, because inflammation is the main cause of the occurrence, development and plaque instability of AS, and the inflammation which causes infiltration of inflammatory cells and release of inflammatory mediators, similar to the phenomenon of heat toxicity, it suggest that antipyretic and detoxicating drugs can play a potential role in stabilizing plaques through anti-inflammation. Therefore, "supplementing Qi and nourishing Yin, activating blood circulation and detoxicating, softening hardness and eliminating accumulation" is an important therapy for diabetes AS."HJJF" (consists of Ginseng, Astragalus membranaceus, Ophiopogon japonicus, Cornus officinalis, Rehmannia glutinosa, Rhubarb, turtle shell, peach kernel, danpi, Coptis chinensis, Salvia miltiorrhiza, yam, Schisandra chinensis), in which, ophiopogon root, ginseng, schisandra are Jin Yuan Dynasty doctor Li Gao «Endogenous and Exogenous differentiation theory» Shengmai Powder ingredients, possessing nourishing Yin and tonifying Qi; cornus, rehmannia and yam are Ming Dynasty doctor Zhang Jingyue prescriptions “Zuo Gui pill” ingredients for nourishing kidney, treating true Yin deficiency; turtle shell, rhubarb, cortex moutan and peach kernel are Eastern Han Dynasty medical scientist Zhang Zhongjing's prescriptions
patients with revascularization or non-revascularization, play plaques and dissipating accumulation. Clinical studies have bridge between metabolic abnormalities and inflammation. proinflammatory factors (TNF-α, IL-6) were lowered, and of renin-angiotensin-aldosterone system (RAAS) in diabetic ACS patients, effectively curb inflammation in diabetic ACS found that HJJF can inhibit insulin resistance, regulate a multi-link, multi-channel, multi-target role in overall regulation, improve. clinical effect and ameliorate cardiac function [5, 19].

This study showed that, After successful diabetes modeling, GSH-Px was activated to a certain extent and compensatory increased, but still insufficient to resist oxidative stress induced by high glucose. After drug intervention, FBG and SOD and GSH-Px were significantly decreased, lipid metabolism disorders were corrected (TC, TG, LDL-C contents were decreased, HDL-C were increased), levels of proinflammatory factors (TNF-a, IL-6) were lowered, and of ER apoptotic signaling molecules GRP78 and JNK transcription levels were down-regulated. HJJF treatment group was better than western medicine group, and the effect was more significant with increasing dose.

5. Conclusion

ERS is a common pathway of multiple stressors and a bridge between metabolic abnormalities and inflammation. The pathological states of diabetes mellitus, such as high sugar, high fat, oxidative stress and inflammation, are all related to ERS, which connects metabolic abnormalities and the development of diabetes mellitus, and causes diabetic atherosclerosis, therefor ERS will become a new target for the treatment of diabetes. The therapeutic effect of HJJF with the functions of nourishing Yin, invigorating Qi, activating blood circulation and detoxifying, lies in correcting insulin resistance, correcting lipid metabolism disorder, improving anti-oxidative stress ability, alleviating inflammatory reaction, inhibiting the coupling reaction of "oxidative stress-endoplasmic reticulum stress-inflammatory response-cell apoptosis", and alleviating inflammation reaction of atherosclerotic plaque in diabetes mellitus, which embodies the prevention and treatment characteristics of integrated intervention, dynamic adjustment and multi-target therapy of traditional Chinese medicine.

Acknowledgements

This research is supported by National Natural Science Foundation of China (81460698); Guangxi emphasis research and development program (2017AB45042); Guangxi Natural Science Foundation (2015GXNSFAA139221).

References


