

## Fatty acids Compositions in Male's Gonads of the Red Sea fish *Rhabdosargus sarba* during the spawning season

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**Abstract:** A total of 27 fatty acids (FAs) were identified in testis throughout the spawning season of male *R. sarba*. In male gonad saturated fatty acids (SFA) were the main fatty acid group in total lipid in testis (34%) followed by PUFA (29.1%) and MUFA (11.6%). In all maturation stages SFA were the main fatty acid group in testis (30.4-35.4%). Of individual fatty acid, the major constituents of SFA were Palmitic acid C16:0 (18.5%) and Stearic acid C18:0 (8.5%) in nearly ripe and ripe stages respectively. Oleic acid C18:1 (8.8%) was found to be the main MUFA in ripe stage and Linoleic acid C18:2 (10.8%) was the main PUFA in nearly ripe stage. During spawning and maturation stages there were a significant differences in total SFA and MUFA ( $P < 0.05$ ).

**Keywords:** Sparidae, *Rhabdosargus sarba*, Fish, Fatty Acids, Spawning, Gonads, Red Sea

## 1. Introduction

Fish lipids are generally characterized by possessing large amounts of saturated and unsaturated fatty acids mostly associated with triacylglycerols and minor amounts of phospholipids [1]. Fatty acids (FA) and particularly polyunsaturated fatty acids (PUFA) are functionally essential for normal growth, development and reproduction in fish [2]. Lipids and specifically fatty acids are the preferred source of metabolic energy for growth, reproduction and swimming in fish especially marine fish as evidenced by the very high oil levels (more than 20% of the wet weight) [3]. Recent studies have clearly shown the importance of polyunsaturated fatty acids (PUFAs) nutritional values for human health [4]. The nutritional importance of fish consumption is in great extent associated with the content of polyunsaturated fatty acids especially omega-3 fatty acids ( $\omega$ -3 FAs) and omega-6 fatty acids ( $\omega$ -6FAs) [5]. The aim of the study to examine the fatty acid profiles in the male of *R. sarba* from the Red Sea. It is emphasized that no earlier studies of the fatty acids composition of this species occur.

## 2. Materials & Methods

### 2.1. Fish Samples

Fish samples were obtained monthly from Bangalah market fish in Jeddah, Red Sea from December to March 2011. Fishes were transported to the laboratory in ice aquarium and then the total length, standard length and weight were measured. Fishes were dissected; testes were removed, weighed and thoroughly examined to determine the maturity stage and the gonad index (testis weight / total fish body weight  $\times 100$ ) was calculated. On the basis of collected commercial data and using the maturity scale described by West (1990) we estimated that the spawning fish samples were in the categories III–VI which characterizes *R. sarba* of mature, ripe and spawning condition.

### 2.2. Lipid Extraction

10 ml of concentrated hydrochloric acid were added to 10g of the sample in a conical flask and immersed in boiling water until the sample was dissolved. At this stage the mixture changed to brown or violet in colour and the fat was seen to collect on the surface. The conical was cooled and the fat was extracted by shaking with 30 ml of diethyl ether. The extract was bowled after allowing the layers to separate into a weighed flask. The extraction was repeated three times more and distilled off the solvent then the fat dried at 100°C, cooled and weighed.

### 2.3. Methylation of Lipid

50 mg of lipid was put in a tube, 5 ml of methanolic sulphuric acid (1 ml conc sulphuric acid and 100 ml methanol) was added and 2 ml of benzene, the tube well closed and placed in water bath at 90°C for an hour and half. After cooling, 8 ml water and 5 ml petroleum was added shaken strongly and the ethereal layer was separated in a dry tube, evaporated to dryness.

### 2.4. Gas Chromatographic (GC) Conditions

The fatty acid methyl esters were analyzed using a Hewlett Packard (HP 6890) chromatography, split/splitless injector and flame ionization detector (FID).

Data were analysed using an independent Student t-test and one way ANOVA for significant differences. The level of significance used was  $P < 0.05$ .

## 3. Results

The fatty acid compositions from testis are presented in Table (1 & 2), 27 fatty acids were identified, ranged from C6:0 to C22:6. On average, 34% saturates, 29% polyunsaturates and 12% monounsaturates, approximately were found in the testis of spawning *R. sarba* (Table1).

**Table 1.** Saturated (SFA), Monounsaturated (MUFA) and Polyunsaturated (PUFA) Fatty acids compositions of *R. sarba*.

SFA	Male
C6:0	0.680 ± 0.365
C8:0	1.387 ± 0.255
C10:0	0.173 ± 0.172
C11:0	0.4729 ± 0
C12:0	0.137 ± 0.038
C13:0	2.608 ± 0.923
C14:0	1.506 ± 0.512
C15:0	1.505 ± 0.552
C16:0	16.35 ± 2.1208
C17:0	1.152 ± 0.4546
C18:0	7.520 ± 1.258
C20:0	0.391 ± 0.340
C21:0	
C23:0	
C24:0	
Total saturates	33.88762 *
MUFA	
C14:1	1.3919 ± 0.429
C15:1	0.5736 ± 0.135
C16:1	2.764 ± 0.471
C17:1	0.491 ± 0.282
C18:1	5.339 ± 2.996
C20:1	0.446 ± 0.126
C22:1	0.6317 ± 0
Total monoenes	11.63913
PUFA	
C18:3	0.360 ± 0.258
C18:2	9.467 ± 0.971
C20:5	3.479 ± 1.049
C20:4	3.384 ± 1.582
C20:3	0.344 ± 0
C20:2	1.234 ± 0.7439
C22:2	6.925 ± 0.494
C22:6	3.882 ± 2.1570
Total polyenes	29.07818

(\*Values are significantly different ( $P < 0.05$ ))

### 3.1. Saturated Fatty Acids (SFA)

It is obvious that the dominant fatty acids among SFA in males was Palmitic acid (C16:0, 16.4%) followed by Stearic acid (C18:0, 7.5%). The most abundant SFA in herring was Palmitic acid C16:0 (ranging from 13.5 to 18.5% in different maturity stages). The relatively high amount of C16:0 present in the nearly ripe stage. However, a decreasing trend in the amount of C16:0 in the testis was noticed during spawning stage (Table2).

**Table 2.** Fatty acid composition in *R. sarba* testes, during them spawning season (2010-2011).

Lipid parameter	Stages			
	Nearly ripe December	Ripe January	Spawning February	Spawning March
Total lipid	0.19	0.1	0.1	0.43
SFA				
C6:0	0.890	1.0662	0.257	0.507
C8:0	1.2307	1.7327	1.1621	1.424
C10:0	0.0849	0.4279	0.1244	0.055
C11:0				0.4729
C12:0	0.1168	0.126	0.195	0.1132
C13:0	1.913	2.217	3.967	2.336
C14:0	1.206	1.1257	2.247	1.446
C15:0	1.1005	1.167	2.301	1.4516
C16:0	18.4915	17.3157	13.541	16.051
C17:0	1.6427	1.4320	0.717	0.8167
C18:0	8.5026	7.279	5.835	8.4645
C20:0	0.24009	0.781		0.1540
Total	35.419*	34.672	30.350	33.296
MUFA				
C14:1	1.083	1.2019	2.027	1.255
C15:1	0.520	0.5486	0.768	0.457
C16:1	3.185	3.1594	2.380	2.332
C17:1	0.2516	0.242	0.731	0.739
C18:1	3.424	3.800		8.792
C20:1	0.357			0.536
C22:1	0.6317			
Total	9.4552	8.952	5.907	14.113
PUFA				
C18:3	0.6149	0.551	0.1406	0.135
C18:2	10.841	8.629	9.419	8.978
C20:5	2.8155	4.530	2.369	4.2037
C20:4	1.788	4.604	2.266	4.876
C20:3	0.3446			
C20:2	2.0917		0.763	0.846
C22:2	7.221	6.863	6.253	7.363
C22:6	2.967	2.863	7.1089	2.589
Total	28.686	28.0416	28.322	28.994

(\* Values are significantly different ( $P < 0.05$ ))

### 3.2. Monounsaturated Fatty Acids (MUFA)

Monounsaturated fatty acids constituted nearly 16% of the total fatty acids in testis of *R. sarba* (Table1). The major MUFA were Oleic acid (C18:1, 5.3%) and Palmitoleic acid (C16:1, 2.8%). MUFA dominated in spawning stage due to the abundance of Oleic acid C18:1 which occurred at more than 30% in this stage. The proportion of C18:1 was three-times higher in the spawning stage (8.8%) and about 2.5 times greater compared to those from nearly and ripe stages (Table2).

### 3.3. Polyunsaturated Fatty Acids (PUFA)

Among PUFAs, Linoleic (C18:2, 9.5%), and (C22:2, 7%) were the major acids. The testis of *R. sarba* had similar PUFA proportions (28%) in the four maturity stages (Table2). The Linoleic acid C18:2 is dominant PUFAs in all maturity stages, the highest proportion of C18:2 occurred in the nearly ripe stage (11%) while the ripe stage contained a relatively lower proportion (9%). Moreover, a relatively higher PUFAs of Docosadienoic acid C22:2 occurred in all maturity stages with an average of 7% (Table2). Highly unsaturated fatty acids (HUFA) namely EPA and DHA are not dominant PUFAs in the testis of *R. sarba*. DHA occurred in greater proportion than EPA in spawning stage spawning (7%) while the other three stages contained a similar proportion (Table2). There were a significant differences ( $P < 0.05$ ) in total SFA and MUFA during spawning season.

Table 2 represents fatty acid (FA) profile of testis samples obtained in the spawning season (December, January, February and March.). No significant variations were identified in the total saturated fatty acid ( $\Sigma$ SFA), monounsaturated fatty acid ( $\Sigma$ MUFA) and polyunsaturated fatty acid ( $\Sigma$ PUFA) amongst months and the highest values were 35.41% (in nearly ripe stage), 14.11% and 29% (at spawning) respectively. During the spawning season (December-March) the highest value for  $\Sigma$ SFA was observed in nearly ripe stage (December) as 35.41% ( $P > 0.05$ ). Palmitic acid (C16:0) and Stearic acid (C18:0) were the major constituents of the total saturated fatty acids. Variations amongst months were also observed for  $\Sigma$ MUFA ranging from 5.9% to 14.11%. Their concentration decreased in both ripe and spawning stage reaching to the lowest value (5.9%) in spawning stage. Oleic acid (C18:1) was found to be the main MUFA varying between 3.4% and 8.8% with the highest value at the end of spawning period (March) followed by Palmitoleic acid (C16:1) where both nearly ripe and ripe stages constituted the highest level (3.2%), whereas the other MUFAs were noticed to be in negligible amount. Significant variations were identified between total saturated fatty acid and monounsaturated fatty acid among months. Fluctuation in  $\Sigma$ PUFA, the testes samples were observed to be constant about 28%. It is clear that Linoleic acid C18:2 was the main PUFA and its level decreased from 10.8% in nearly ripe stage to 8.6% in ripe stage. Docosadienoic C22:2 is found to be the second major PUFA, spawning and nearly ripe testis had the highest level among months (7.4%, 7.2%) respectively.

## 4. Discussion

There is number of classifications available by which fish is divided into groups, according to their lipid content. The species *R. sarba* belong to the group of high protein-low fat category. Saturated fatty acids (SFA) of total lipid constituted the majority of the fatty acids pool followed by polyunsaturated fatty acids (PUFA) and monounsaturated

fatty acid (MUFA) in males of *R. sarba*. The results are in agreement with those reported by [6-8] for sea bass (*D. labrax*).

The major fatty acids in male of *R. sarba* were Palmitic acid (C16:0), Stearic acid (C18:0), Oleic acid (C18:1), Linoleic acid (C18:2), Eicosapentaenoic acid, (EPA, C20:5), Arachidonic acid (AA, 20:4), C22:2 and Docosahexaenoic acid (DHA, C22:6). This result is similar to those reported for some selected Indian Fishes [9]. Grigorakis *et al.* [10] also determined that these fatty acids constituted the basic components of fatty acids of gilthead sea bream (*S. aurata*). Similarly, Varljen *et al.* [11] found that these fatty acids were the basic fatty acids of *Diplodus vulgaris*. Also, the same results were recorded for *S. aurata* and *D. sargus* by Özyurt *et al.* [12]. The C16:0 and C18:1 fatty acids were mainly catabolic for energetic purposes [13]. High amounts of all these acids were consumed during fish growth and development and they were easily catabolic by the mitochondria [14].

The saturated fraction constituted in male 33.88% of the total fatty acids, Palmitic acid C16:0 is the most important fatty acid followed by Stearic acid C18:0. The dominance of Palmitic acid in fish lipid has been reported by other authors [15-18]. In the present study the Palmitic acid showed higher level in testes which is noticed for being a predominant source of potential metabolic energy in fish during growth [19]. Similar result has been reported previously in the roe of herring [20]. Also, this may be indicator for male need more supplying of energy requirements for changing the sexes in *R. sarba* from male to female (hermaphrodite).

As far as for MUFA fraction Oleic acid (C18:1) was the basic fatty acid. Similar results were reported for gilthead sea bream by Grigorakis *et al.* [10], and for sea bass by Bhoury *et al.*, [21]. In the present study Linoleic acids (C18:2) was the prominent PUFA (9.47%), this result is in accordance with Bayir *et al.* [22] who reported that C18:2 was the major PUFA for gilthead sea bream and two-banded bream (Sparidae). Similar finding have been reported in three Tunisian silverside fish [23]. Higher proportion of C18:2 have been reported in Baltic herring which have been linked to dietary factors [24]. The results concerning variation in the present study generally agree with the previously reported finding for sparidae species where fatty acid compositions studied in relation to the reproductive cycles, specially reduction in PUFA including EPA and DHA [25].

Marine fish have low or no capacity to synthesize highly unsaturated fatty acids (HUFAs) from C18 fatty acids. HUFAs are important components of cell membranes and are thought to play important roles in membrane fluidity, modulation of enzyme activity, neural development and regulation of stress resistance. Especially, Eicosapentaenoic acid (EPA: 20:5n-3) and Docosahexaenoic acid (DHA: 22:6n-3) were considered as dietary essential fatty acids for normal growth and survival in most marine fish [26]. Linoleic acids (C18:2) was the prominent PUFA in males *R.*

*sarba* followed by C22:2 (6.93%), Docosahexaenoic acid (DHA) Eicosapentaenoic acid (EPA) and Arachidonic acid (AA). This result is similar to those reported for Malaysian marine fishes, Eicosapentaenoic acid (EPA) and Docosahexaenoic acid (DHA) were found more in marine fishes PUFAs than in freshwater fishes [27].

In the present study, arachidonic acid (AA) showed lower levels (3.4%). Marine fish species generally have only PUFA which originate from marine phytoplankton. Therefore AA levels were generally low or negligible in total fatty acids of lipid of marine fish species [28], the content of AA in marine fishes were lower than freshwater fishes. Therefore, high levels of AA may be useful as a lipid biomarker of herbivorous fishes which prefer seaweed [29].

Although C20:4 has similar biological importance as EPA and DHA, it is often neglected in fish because it occurs only at very low concentration, despite possessing vital functions as main precursor of various Eicosanoids [3], that have important roles in a variety of physiological functions including osmoregulation, cardiovascular function and the function of reproductive systems [30]. Prostaglandins (PG) plays an important role in fish reproduction [31] with AA being the principle PG precursor involved in spawning activity of fishes including sperm production [32] and its lower levels probably due to its mobilization. Evidence for the importance of AA in reproduction was first identified in European sea bass brood stock [33,34].

DHA content of lipids in tropical and subtropical marine fish species were reported to be lower than those of arctic and subarctic ones [35]. Fishes living in warm-water seas do not appear to require as much DHA in their cell membranes as cold-water species [36]. The lipid content of tropical fish species is generally influenced by high ambient temperature and the membrane lipids of these fishes are easily fluidized even if the major fatty acids in their lipids are composed of saturated and monoenoic fatty acids.

Levels of C16:0 and C18:1 and low levels of HUFA were characteristic for neutral lipids of most marine fish [37]. This is also in concordance with the common opinion that fish species accumulate depot lipids composed mainly of saturated and monoene fatty acids [38].

## 5. Conclusions

From the present study it has been shown that 27 fatty acids were detected in testis of male *R. sarba*. Saturated fatty acids (SFA) were the main fatty acid group in total lipid followed by polyunsaturated fatty acids (PUFA) and monounsaturated fatty acids (MUFA). The major constituents of SFA were palmitic and stearic acids in nearly ripe and ripe stages respectively. Oleic acid was found to be the main MUFA in ripe stage and linoleic acid was the main PUFA in nearly ripe stage.

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