Predictive role of αvβ3 integrin, and VEGF in colorectal adenocarcinoma progression in Iraqi patients

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Abstract: Angiogenesis, the formation of new blood vessels from pre-existing ones, is enhanced in various pathological conditions including rheumatoid arthritis, diabetic retinopathy, and cancer development. Angiogenic processes are regulated by both growth factors, such as vascular endothelial growth factor (VEGF), and adhesion molecules, such as integrin. Understanding the molecular mechanisms that underlie regulation of angiogenesis especially VEGF function, is essential for the development of safe and effective antiangiogenic therapies. Thus, current study was aimed to investigate predictive role of β3 integrin and vascular endothelial growth factor (VEGF) protein expression in colorectal adenocarcinoma sample from Iraqi patients, through linking its expression with tumor histopathological variables (stage, grade, grade, and lymph node involvement), by using Immunohistochemical staining method. Study done on 35 colorectal cancer samples and their respective resection margins. Present study demonstrated that, the positive expression rate of integrin β3 and VEGF in non-tumor colorectal mucosa (25.3333 ± 1.974842, and 32.4 ± 1.974842) was significantly lower than that of the colorectal cancer (CRC) tissue (76.47059 ± 2.878562, and 79.45714 ± 2.293705; P < 0.05). Moreover, when CRC samples breakdown according to histopathological variables, In patients of stage C-D, poorly differentiated, and with lymph node (L.N) invasion, the positive expression rates of integrin β3 were significantly higher than those in patients of patients with stage A,B, well or moderately differentiated, and without lymphatic metastasis (P < 0.05, P < 0.05, and P < 0.05; P < 0.05, P < 0.05, and P < 0.05), respectively. In conclusion, Integrin β3 and VEGF expression can synergistically enhance tumor angiogenesis, and may play a crucial role in invasion and metastasis of colorectal carcinoma. Therefore, they may be prognostic biomarkers and novel molecular therapeutic targets.

Keyword: Colorectal Cancer, Angiogenesis, VEGF

1. Introduction

Angiogenesis is the formation of new capillaries from preexisting vessels, it is essential for several normal physiological events such as embryonic vascular development, wound healing, and reproduction. It is also involved in pathological processes, including tumor growth, diabetic microvascular disease, and rheumatoid arthritis [1, 2]. Angiogenesis depends on input from growth factors and vascular cell adhesion signals [3, 4]. Among angiogenic growth factors and adhesion molecules, are vascular endothelial cell growth factor-A (VEGF-A) and αvβ3 integrin, respectively, are two well-established representatives. To date, VEGF-A is the only growth factor proven to be specific and critical for blood vessel formation [5, 6]. Integrin are transmembrane glycoproteins composed of non-covalently associated α- and β-chains which recognize proteins of the extracellular matrix (ECM). The importance of αvβ3 in angiogenesis has been underscored based on the findings that αvβ3 is prominently expressed on the surfaces of endothelial cells and is highly upregulated on angiogenic blood vessels such as those in solid tumors and in granulation tissue at the base of healing wounds [7]. Recent investigations have shown that high expression of integrin β3 is positively correlated with invasion and metastasis of cancer cells and tumor angiogenesis [8, 9], but only a few studies have investigated the relationship between integrin β3 and
prognosis of colon cancer. The molecular mechanisms by which colorectal cancer cells exploit the hepatic microenvironment for selective growth and survival remain obscure. The report by Conti et al. has now identified essential functions for αvβ3 integrin in promoting metastatic colorectal adenocarcinoma cell growth and survival in the liver [10]. In our study, we aimed to evaluate integrin β3 and vascular endothelial growth factor (VEGF) protein expression in colorectal adenocarcinoma (CRC). And its relationship to pathological parameters such as tumor stage, grade, and lymph node infiltration.

2. Materials and Methods

2.1. Patients and Sampling

Thirty five patients with colorectal adenocarcinoma, who were confirmed histopathologically, were included in this study. Their age were ranged from 20-80 years. Paraffin embedded blocks of tumor and resection margins were retrieved along with the histopathological report of each patient from histopathological laboratory. For staging of the cancer, astler-coller staging system was adopted in this study [11]. In addition, resection margins were confirmed again to be free of malignancy. Adequate thin paraffin embedded sections (5µm thick) of tumor and resection margins were prepared on positively charged resection margins, based on t.Test

2.2. Immunohistochemical Detection of αvβ3 Integrin, and VEGF

A primary monoclonal antibody against "αvβ3 integrin, and VEGF" (US biological, USA) reacts with their antigen. Biotinylated secondary antibody then reacts with the primary antibody. This is followed by the attachment of an enzyme-conjugated streptavidin to the biotins on the secondary antibody. The enzyme converts a substrate to a colored reaction product. And the procedure has been done according to manufacturer instructions (US biological, USA). There after slides were examined by histopathologist under light microscope (100x).

3. Results

3.1. Histopathological Data

Thirty five patients with colorectal adenocarcinoma were investigated. According to the histological differentiation, tumors were broken down in to three groups, well differentiated (WD, n=7), moderately differentiated (MD, n=20), and poorly differentiated (PD, n=8), moreover, patients were further grouped according to their histopathological criteria, as follow: tumor stage (A, n=6, B, n=10, C, n=11, and D, n=8), lymph node involvement (free, n=16, and involved, n=19).

3.2. Tumor Sites versus Resection Margins

Based on immunohistochemical staining method, and according to statistical analysis there were significant difference based on mean ± standard error (SE) of both αvβ3 integrin, and VEGF protein expression between tumor sites and their resection margins (p<0.001 and, 79.45714±2.293705, 32.4± 1.974842; p=0.001 respectively), (table 1). A noteworthy here is that high intensity of staining is found in the periphery of necrotic foci of the colorectal cancer, labeling with the VEGF antibody distinctly intensified in tumor epithelium adjacent to the necrotic foci. Moreover, the tumor stroma cells also labeled for VEGF, labeled stromal cells included fibroblasts and smooth muscle cells. αvβ3 and VEGF IHC staining in tumor sample and resection margin are shown in figure 1.

<table>
<thead>
<tr>
<th>Sample</th>
<th>No.</th>
<th>αvβ3 Mean ± SE</th>
<th>VEGF Mean ± SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>35</td>
<td>25.3333±1.974842</td>
<td>32.4± 1.974842</td>
</tr>
<tr>
<td>Tumor</td>
<td>35</td>
<td>76.47059±2.878562</td>
<td>79.45714±2.293705</td>
</tr>
</tbody>
</table>

* (P<0.005).

3.3. Correlation among Protein Expression of αvβ3 and VEGF with Different Histopathological Variables

αvβ3 and VEGF protein expression in colorectal adenocarcinoma were analyzed against the different histopathological features of the tumors based on Spearman’s correlation. As shown in Table 2, and 3, current study demonstrated that there were significant correlations between both αvβ3 and VEGF expression with tumor stage, and grade depending on mean expression ±SE value (P<0.05, P<0.05 respectively). Moreover, a significant differences were found when we compare the expression level of both markers according to L.N involvement (P<0.05, P<0.05 respectively) table 4.

<table>
<thead>
<tr>
<th>Stage</th>
<th>No.</th>
<th>Mean ± SE αvβ3</th>
<th>VEGF</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>6</td>
<td>51.33 ± 2.33</td>
<td>59.00 ± 4.80</td>
</tr>
<tr>
<td>B</td>
<td>10</td>
<td>67.60 ± 2.67</td>
<td>75.00 ± 2.15</td>
</tr>
<tr>
<td>C</td>
<td>11</td>
<td>85.45 ± 2.01</td>
<td>84.27 ± 1.82</td>
</tr>
<tr>
<td>D</td>
<td>8</td>
<td>96.50 ± 0.68</td>
<td>93.75 ± 1.84</td>
</tr>
<tr>
<td>LSD value</td>
<td>---</td>
<td>6.428 *</td>
<td>7.334 *</td>
</tr>
<tr>
<td>* (P&lt;0.05).</td>
<td></td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>Grade</th>
<th>No.</th>
<th>Mean ± SE αvβ3</th>
<th>VEGF</th>
</tr>
</thead>
<tbody>
<tr>
<td>WD</td>
<td>6</td>
<td>52.80 ± 4.57</td>
<td>60.67 ± 5.53</td>
</tr>
<tr>
<td>MD</td>
<td>19</td>
<td>74.47 ± 3.18</td>
<td>78.26 ± 1.76</td>
</tr>
<tr>
<td>PD</td>
<td>10</td>
<td>92.85 ± 2.84</td>
<td>93.00 ± 1.80</td>
</tr>
<tr>
<td>LSD value</td>
<td>---</td>
<td>12.935 *</td>
<td>7.934 *</td>
</tr>
<tr>
<td>* (P&lt;0.05).</td>
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</table>
Table 4. Expression pattern of αvβ3 and VEGF along with L.N involvement of CRC, based on spearman’s correlation (r_s)

<table>
<thead>
<tr>
<th>LN</th>
<th>No.</th>
<th>Mean ± SE αvβ3</th>
<th>VEGF</th>
</tr>
</thead>
<tbody>
<tr>
<td>Free</td>
<td>16</td>
<td>64.90 ± 3.85</td>
<td>69.00 ± 2.93</td>
</tr>
<tr>
<td>Involved</td>
<td>19</td>
<td>90.11 ± 1.74</td>
<td>88.26 ± 1.68</td>
</tr>
<tr>
<td>LSD value</td>
<td>----</td>
<td>10.391 *</td>
<td>5.066 *</td>
</tr>
</tbody>
</table>

* (P<0.05).

Figure 1. Immunohistochemical staining of αvβ3 and VEGF in colorectal adenocarcinoma section and their resection margins by DAP (brown color) counterstained with hematoxylin. (A) αvβ3 protein expression in tumor cells. (B) Resection margin stained with αvβ3 integrin. (C) VEGF expression in tumor section. (D) Resection margin stained with VEGF. Magnification power (400X).

4. Discussion

Since, current clinical methods can not accurately predict which patients will develop metastasis. Thus, to develop effective new strategies for the prediction, diagnosis and treatment of cancer patients, the molecular mechanisms controlling tumor aggressiveness must be identified. Therefore, Integrin and VEGF could be one of such mechanisms. Integrin β3 may enable VEGF to stimulate endothelial cell proliferation and capillary angiogenesis through activating VEGF receptor-2 (Flk-1) and VEGF receptor-3 (Flt-4) [12, 13]. Current study had demonstrated a significant over expression of αvβ3, and VEGF protein expression in tumor tissue in compares with their resection margins (p<0.05, and p<0.05 respectively, table 1). This observation came in compatible with previous studies that focused on VEGF in colorectal and other tumors since; they illustrated a significant increase in VEGF protein expression with respect to tumor status, nodal metastasis, and with poor clinical outcome[14]. Moreover, current results solidify previously published data which mentioned that αvβ3 is positively correlated with angiogenic activity, invasion, and metastasis, [15]thus, it consider the leading edge of liver metastasis. Since, hypoxia is an important stimulus for mediating VEGF mRNA cellular expression during angiogenic response [16, 17] and the current data demonstrated an increase of VEGF expression around necrotic tissue. Thus, it would be acceptable to anticipate
that hypoxia could have the upper hand behind this current outcome. Moreover, current study showed that integrin β3 and VEGF protein expression rate were significantly associated with tumor stage, poor differentiation, and lymphoid node invasion, (P< 0.05, P< 0.05& P< 0.05 respectively; able 2, 3, & 4). Thus, it came with previous studies; they mentioned that an over expression of β3 integrin generally appears to be positively correlated with tumorigenicity. For example, expression of the β3 integrin subunit in melanoma in situ has been found to correlate with tumor thickness, the ability to invade and metastasize, and poor prognosis [18]. Also, increased expression of integrin β3 in gastric cancer influenced the adhesion between tumor cells and ECM. β3 integrin may influence signal transduction, thereby changing the biological behavior of tumor cells, and enhancing the potency of infiltration and migration [19]. Since, angiogenesis not only accelerates tumor growth, but also increases the opportunity of tumor cells for invading the vasculature, hence it promotes tumor metastasis. Thus, that could explain higher VEGF expression at the invasive tumor edge and strong association with LN metastasis. As the bridge between vascular endothelial cells and ECM, integrin play an important role in angiogenesis [20]. Previous studies have shown that integrin αvβ3 is minimally expressed on resting or normal blood vessels, but is significantly up-regulated in vascular cells within human tumors, and has been implicated in tumor-induced angiogenesis [21, 22]. Unligated integrin can act as a negative regulator of cell survival, initiating a process of “integrin mediated death” [23]. In our assay system, β3 integrin are presumably in an activated state because of VEGF-A released from the tumor cells. Evidence suggests that integrin may associate with a number of immobilized pro-angiogenic molecules. β3 integrin may interact with distinct isoforms of VEGF-A and these interactions may enhance endothelial cell adhesion, migration and survival [24, 25]. In addition to interactions with secreted and matrix bound growth factors, integrin also associate with growth factor receptors. VEGF-A-mediated activation of VEGFR2 recruits Src and facilitates phosphorylation of the β3 cytoplasmic tail [26]. The phosphorylated β3 tail may promote interactions between αvβ3 and VEGFR2, thereby enhancing endothelial proliferation and survival [26]. Finally, further studies based on pre-malignant, and linking expression level of VEGF with microvascular density are needed to improve current results.

5. Conclusion

In summary, integrin β3 and VEGF are consideringly a guide marker which synergistically enhance tumor angiogenesis, and may play crucial roles in invasion and metastasis of CRC. Therefore, they can be used as biomarkers for diagnosis and prognosis, and as novel molecular therapeutic targets.

References


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