

Nitrate anion as a probe for electrostatic interactions in complexes protein-ligand

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Abstract: We are proposing a new technique for studying interaction of charged biomolecules. It is based on a difference between nitrate and chloride anion influence on electrostatic interactions in order to detect the role of positively charged guanidine groups of proteins. This comparison was demonstrated for human serum albumin (HSA) interaction with a fluorescent reporter, CAPIDAN. Nitrate anions lower their binding constant (K). There are two causes that are responsible for that K decrease induced by nitrate. The first, rise of the ionic strength (like to chloride), and the second, a specific one: at the same concentration and ionic strength nitrate anion more significantly lowers K than chloride anion. The study of electric conductivity of chloride and nitrate salts shows that nitrate anions better than chloride anions form complexes with guanidine cations. Therefore it can be assumed that the nitrate-chloride techniques detect a direct contact of negatively charged carboxyl of CAPIDAN with positively charged arginine residues of HSA. HSA site I includes Arg 218 and Arg 222. It is possible that the CAPIDAN carboxyl binds to one of these arginines.

Keywords: Human Serum Albumin, Drug-Binding Site, Negatively Charged Ligands, Nitrate Anion Blocking Arginine Charge, Fluorescent Probe CAPIDAN

1. Introduction

Human serum albumin (HSA) transports many metabolites. To perform that function, special pockets (binding sites) exist in HSA molecules. Besides metabolites, these sites can bind xenobiotics including drugs as well as aromatic dyes [1,2]. Therefore such dyes, especially fluorescent ones, are widely used for the binding site study.

Recently one of the dyes, named CAPIDAN (Fig.1) [3], has been used to detect HSA molecule changes at some diseases, and some diagnostically important data have been obtained [4]. To clarify the nature of these changes in HSA binding sites, nitrate anions have been used: there is some data that nitrate is able to quench dynamically protein fluorescence and therefore it can be used for studying accessibility of protein fluorophores to water [5, 6].

We have tried to use the property of nitrate for testing HSA-bound CAPIDAN accessibility to water in pathologi-

cal processes. Adding nitrate anion as well as chloride substitution by nitrate led to decrease of CAPIDAN fluorescence intensity [7, 8]. However this effect is different in normal sera and at some psychic abnormalities [7, 8]. This

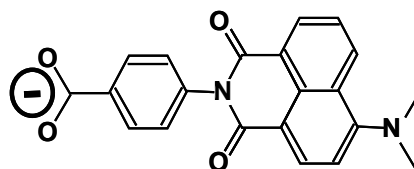


Figure 1. Fluorescent probe CAPIDAN, negatively charged at neutral pH.

study is an attempt to clarify mechanism of the fluorescence quenching by nitrate anions in the system fluorescent probe-HSA. Results that be obtained are found to be unexpected.

2. Methods

Fluorescent probe CAPIDAN, N-(p-carboxyphenyl) imide of 4-(dimethylamino) naphthalic acid, has been synthesized and was kindly provided by B.M. Krasovitsky and colleagues (Institute of Monocrystals, Kharkov, Ukraine).

Lyophilized powder of human serum albumin, fatty acid free, was purchased from Sigma-Aldrich (cat. A 1887). It was solved in a buffer, 0.01 mol/litre sodium phosphate + 2 mmol/litre EDTA pH 7.4. This buffer was a base for all solutions used in fluorescence measurements. Guanidine hydrochloride and other salts were obtained from the same source.

Fluorescence spectra were measured using Hitachi F 4000 spectrofluorometer. Fluorescence decay was measured using a Pico-Quant instrument; the excitation source was a pulse diode with spectral maximum of 455 nm; the Instrument Response Function, IRF, was close to 0.6 ns. CAPIDAN fluorescence decay was approximated as a sum of exponentials using the chi-squared criterion.

Conductivity of salt solutions was measured at 250 C using conductometer InoLab Cond 720.

3. Results

CAPIDAN fluorescence intensity in water is very low, however it rises many times in the presence of HSA as a result of CAPIDAN binding to HSA [3, 4]. Increasing ionic strength leads to fluorescence decrease (Fig.2). Probably

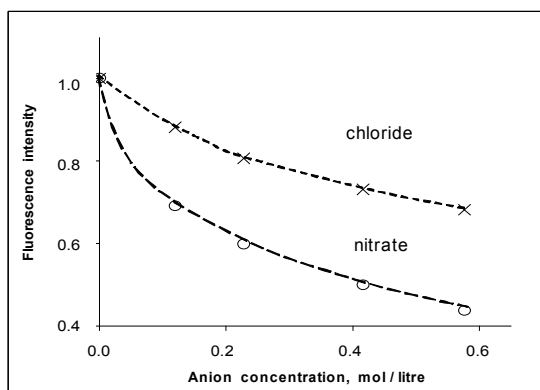


Figure 2. Lowering CAPIDAN fluorescence intensity in HSA solutions due to KCl or KNO₃ addition. HSA 30 μmol/litre, CAPIDAN 14 μmol/litre.

the ionic strength rise suppresses Coulomb interaction of the CAPIDAN negative charge with positive charges of the HSA binding site [3] – like to other negatively charged aromatic ligands [1,2] (some recent similar examples are [9,10,11] etc.).

Ionic strength risen by NaCl or KCl addition gives the same effect: i.e. cations Na⁺ and K⁺ are equivalent. Meanwhile nitrate anion suppresses the CAPIDAN fluorescence more effectively than chloride. So, two effects exist: an unspecific effect of the ionic strength rise and a specific action of nitrate on CAPIDAN fluorescence. That phenomenon is seen also in Fig.3 where chloride anions were

substituted equimolarly by nitrate at invariable ionic strength.

What is the cause of that difference between chloride and nitrate effects?

3.1. A Hypothesis of the Dynamic Quenching

Nitrate anions are able to quench dynamically the intrinsic protein fluorescence leading to decrease of fluorescence decay time τ [6]. Stern-Volmer description of a simple dynamic quenching [12] is

$$1/\tau = 1/\tau_0 + k_q[Q]; \tau_0/\tau = 1 + \tau_0 k_q[Q] \quad (1)$$

where τ_0 and τ are the decay time in the absence and the presence of a quencher, respectively; $[Q]$ is the quencher concentration; k_q is the rate constant of effective quencher-chromophore collisions. If the steady-state fluorescence intensity F is proportional to τ , then a common equation originates from (1) [13]:

$$F_0/F = 1 + K_{SV}[Q] \quad (2)$$

However the data of Fig. 2 cannot be described by the simple (2) (not shown). So, the existence of a simple dynamic quenching is not obvious. A more complex quenching can not be excluded on this step of the study because the initial CAPIDAN fluorescence decay in HSA has a complex nature [3, 4, 14, 15]. Different CAPIDAN molecules in HSA have different environment producing three species of bound probe with three types of decay:

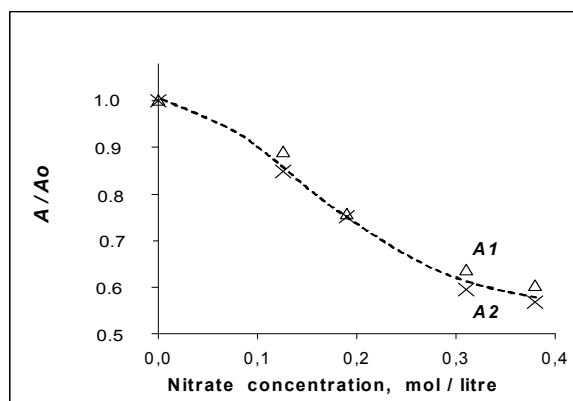


Figure 3. Lowering amplitudes of HSA-bound CAPIDAN fluorescence decay components due to equimolar substitution of chloride anions by nitrate. Summary concentration of chloride and nitrate was 0.63 mol/litre. A_0 is the amplitude without nitrate.

$$F(t) = A_1 \exp(-t/\tau_1) + A_2 \exp(-t/\tau_2) + A_3 \exp(-t/\tau_3) \quad (3)$$

where t is time after excitation. Amplitudes A_1 , A_2 and A_3 are proportional to populations of these three species. τ_1 value is close to 9.7 ns, τ_2 is near 3.6 ns and τ_3 is about 1 ns (Table 1). Two first species are responsible for about 90-95% (40-50 per cent each) of total fluorescence of HSA-bound CAPIDAN. Ionic strength rise does not influence decay times τ_i and decreases all three amplitudes A_i ; i.e. CAPIDAN binding to HSA decreases.

Dynamic quenching should influence these τ_i . If nitrate dynamically quenches CAPIDAN fluorescence in HSA, then decay times should decrease in accordance with the Stern-Volmer theory (1). Meanwhile substitution of chloride by nitrate does not change any τ (Fig.4 and Table 1). So, the major sign of dynamic quenching, i.e. τ decrease, is absent. Nitrate lowers amplitudes A_i . The most probable explanation of this phenomenon is suppressing CAPIDAN binding to HSA induced by nitrate. However nitrate does it significantly more effectively than chloride.

3.2. Suppressing CAPIDAN-Albumin Binding Caused by Nitrate

Nitrate influence on the CAPIDAN binding to HSA was tested using HSA titration by CAPIDAN. Data on A_1 changes are presented in Fig.5 as a double reciprocal graph proposed by Klotz [2]. A_1 is the amplitude of the first component of (3); it is proportional to number of the first species molecules bound to HSA and having $\tau_1 \approx 9.7$ ns. Reciprocal value of CAPIDAN concentration is shown on horizontal axis. The experimental data can be fitted to a straight line. This line has an intercept on vertical axis equal to $1/A_{1max}$ where A_{1max} is proportional to number of HSA binding sites for the first CAPIDAN species. The intercept on the horizontal axis is approximately equal to the binding constant K_1 for the species. As it was shown, even partial substitution of chloride by nitrate decreases K_1 by 2.9 times. In the case of CAPIDAN second species ($\tau_2 \approx 3.6$ ns) K_2 decreases by 1.8 times (not shown). That substitution chloride/nitrate does not influence significantly number of binding sites (vertical intercepts); this is also a sign of absence of statically quenched CAPIDAN molecules.

Table 1. Parameters of CAPIDAN fluorescence decay in HSA solution as a function of the ionic strength at NaCl concentration rise. Amplitudes A_i are divided by their magnitudes at the lowest ionic strength 0.02 mol/litre. Decay time τ_i error is ± 0.1 ns.

Ionic strength, mol/L	0.02	0.16	0.52
$A_1 / A_1(0.02)$	1.00	0.52	0.32
$A_2 / A_2(0.02)$	1.00	0.63	0.44
$A_3 / A_3(0.02)$	1.00	0.80	0.62
τ_1 , ns	9.7	9.7	9.8
τ_2 , ns	3.9	3.6	3.6
τ_3 , ns	1.1	1.0	1.0

These data shows that chloride and nitrate anions suppress CAPIDAN binding to HSA but nitrate is significantly more effective at the same concentration. Nitrate lowers the binding constant K and does not influence number of binding sites.

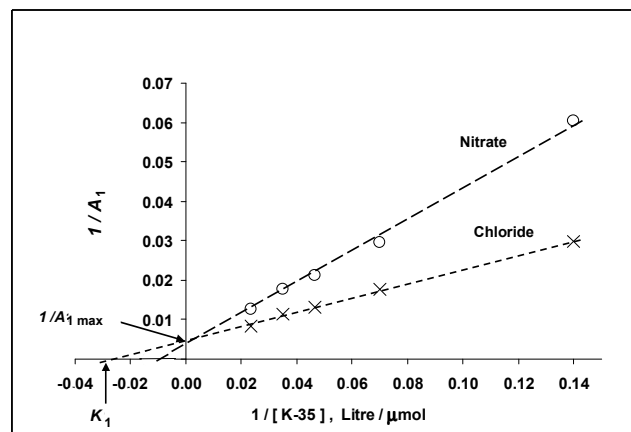


Figure 5. Titration of HSA by CAPIDAN in the presence of chloride 0.63 mol/litre and after equimolar substitution of 0.38 mol/litre chloride by nitrate. A_1 is amplitude of the CAPIDAN first species, A_{1max} is its maximal value. K_1 is the binding constant.

3.3. Some Physical Chemical Differences Between Chloride and Nitrate Anions

Why is nitrate a more effective suppressor of CAPIDAN binding than chloride? CAPIDAN binds with the first drug-binding site [3,4,14]. Negative charge of ligands promotes their binding to HSA [1,2,10,11]. Crystallographic data have shown [16,17,18] that a cluster of positively charged lysine and arginine residues is localized on the boundary site I/water (Fig.6). Probably CAPIDAN negative charge interacts with this cluster; this hypothesis can explain the ionic strength effect (Fig.2 and Table 1) but does not clarify nitrate/chloride difference.

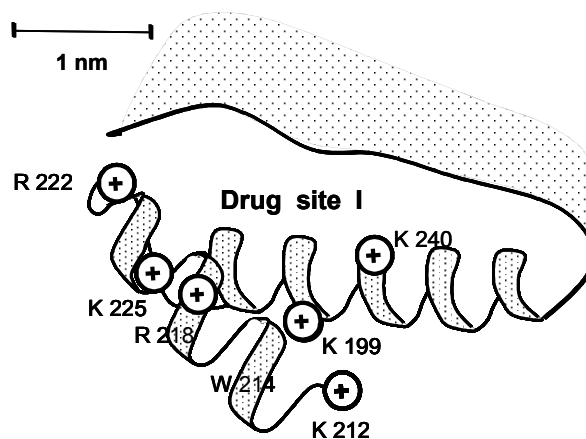


Figure 6. Schematic representation of entrance into the albumin first drug-binding site on the base of crystallographic data [16; 17; 18]. The single tryptophan residue (W) and positively charged residues of lysine (K) and arginine (R) are shown.

It can be supposed that nitrate interacts with these HSA charges more strongly than chloride. Maybe, that interaction occurs with lysine ammonium groups, as it has been proposed by Klotz [1]. Indeed, as is known, there exists a significant difference between interaction of nitrate and

chloride with ammonium: e.g. solubility of ammonium nitrate in water is 3.4 times more than ammonium chloride [19]. However in the case of CAPIDAN binding there is an inverse situation. At the same time solubility of guanidine nitrate is 6 times lower than guanidine chloride [19,20]. Thus, arginine guanidine groups could be assumed as probable targets for nitrate anions.

To obtain proof for this hypothesis, electric conductivity of guanidine salt solutions have been measured.

3.4. Conductivity of Guanidine Salt Solutions

Electric conductivity of salt solutions originates from ionic components of these solutions. So, KCl dissociates on K^+ and Cl^- in the range of concentrations used above (see data of Fig.2–5 and Table 1). Conductivity of KCl solutions rises as a linear function of KCl concentration (Fig.7). This is a known fact [19]. Conductivity of guanidine chloride (GuCl) is approximately the same as KCl (Fig.7), i.e. GuCl almost fully dissociates on Gu^+ and Cl^- , and moreover cations Gu^+ have conductivity close to K^+ . Nitrate anions have conductivity close to chloride anions if the cation is K^+ [19]. Therefore in mixtures K^+ , Gu^+ , Cl^- and NO_3^- the total conductivity might be expected to be proportional to the total salt concentration C . Results of conductivity changes due to KCl or KNO_3 addition to 0.4 mol/litre guanidine hydrochloride are presented in Fig.8 where conductivity is normalized on the total salt concentration. C here is close to ones in experiments on CAPIDAN binding to HSA (see above). As can be seen, there is a significant difference between KCl and KNO_3 effects. In mixtures K^+ , Gu^+ and Cl^- this proportionality between conductivity and C is confirmed, and the ratio κ/C is only very slightly dependent on C (Fig.8, upper curve). However it is not so in the case of mixing Gu^+ with NO_3^- where the ratio κ/C falls

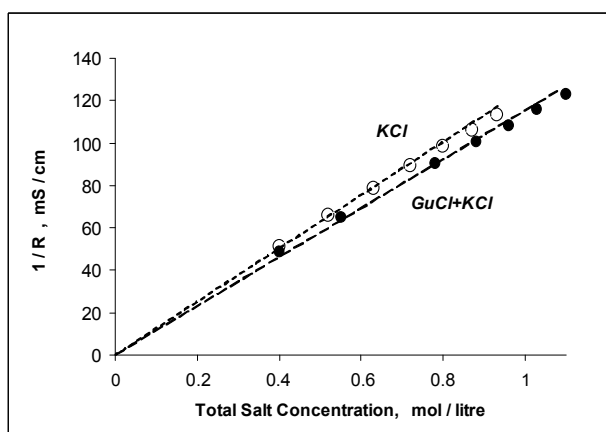


Figure 7. Conductivity of potassium and guanidine hydrochloride solutions. KCl (upper curve) or GuCl (lower curve) were added to 0.4 mol/L KCl solution. R – electric resistance.

significantly with C increasing (Fig.8, lower curve). The single explanation of that decrease is nitrate-guanidine complex formation because complexes anion-cation do not

participate in conductivity.

Thus, conductivity data show that an interaction of nitrate anions with guanidine cations is stronger than chloride interaction with guanidine. As a result, nitrate shields the guanidine positive charge.

4. Discussion

Results presented above show that chloride and nitrate anions suppress binding of negatively charged fluorescent probe CAPIDAN with drug-binding site I of HSA but nitrate suppresses it more effectively. Obviously the binding strengthens due to Coulomb interaction of CAPIDAN charged carboxyl with positive charges of HSA [3]. Those charges can be attributed to amino groups of lysine or guanidine groups of arginine residues. Therefore, the rise of the ionic strength by chloride or nitrate leads to lowering CAPIDAN binding. At the same time, nitrate has an additional, specific effect. Conductivity data show that nitrate anions form uncharged complexes with guanidine cations while chloride with the same concentration almost does not. This fact allows to suggest that nitrate specific effect on CAPIDAN binding is due to shielding arginine charges. There is data on CAPIDAN binding with HSA drug site I [4,14]. In this site of crystallized HSA, a cluster of positive charges including Arg218 and Arg222 exists [16,17,18]. Data of neutron [21] and X-ray small-angle scattering [22] show that size of HSA molecule is similar in crystals and solutions; perhaps, the site I structure is similar as well. So, it can be suggested that negatively charged carboxyl of CAPIDAN binds with Arg218 or Arg222 of the site I, and nitrate suppresses this interaction. Binding of an anionic drug, warfarin, to the same site significantly enhances after Lys199 replacement by an uncharged residue but falls threefold after Arg218 replacement [16,23]. These

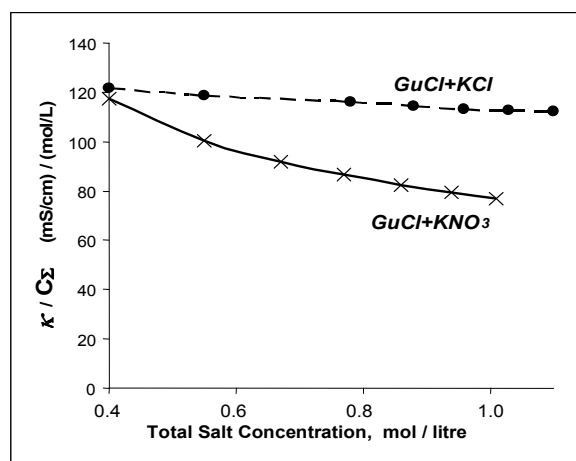


Figure 8. Ratio of the mean molar conductivity κ , and the total salt concentration C_{Σ} of 0.4 mol/litre GuCl water solution after addition KCl (upper curve) or KNO_3 (lower curve).

data support the suggestion that CAPIDAN also interacts with arginines of the site I and nitrate anions hinder this

interaction.

5. Conclusions

Electrostatic interactions of charged amino acid residues play a very important role in formation of complexes protein-protein as well as protein-hormone, drug etc. Meanwhile contribution of different residues into these interactions is unknown very often. The data proposed above shows a way to detect participation of the arginine charge in those interactions. This approach has been tested on complexes of human serum albumin (HSA) with a fluorescent probe as a ligand. This complex is formed due to different forces including electrostatic [3]. Comparing the effects of nitrate and chloride anions on the interaction allows to detect the contribution of arginine residues. So, nitrate anion can be assumed to be a novel probe to study interaction of charged ligands with arginine positive charges of HSA and, probably, other proteins in solutions.

That interaction is significant if the distance between CAPIDAN carboxyl and this HSA guanidine group is rather small, i.e. several Angström. Therefore some small conformational changes of the protein binding site, including pathology-induced, could be detected using nitrate as a probe.

Acknowledgments

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