

Asymptomatic Bacteriuria and Their Antimicrobial Susceptibility Pattern Among Pregnant Women Attending Antenatal Clinics at Mizan Aman Town, Southwestern Ethiopia

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Abstract: *Introduction:* Failure to identify asymptomatic bacteriuria (ASB) is responsible for its improper clinical management. This may lead to severe health complications during pregnancy. However, little is known about antimicrobial susceptibility of ASB in Ethiopian pregnant women. Therefore, the study was conducted to assess the prevalence of ASB and its antimicrobial susceptibility pattern in the target population. *Methods:* A cross-sectional study was conducted during September-December of 2017 at Mizan-Aman town, south-western Ethiopia among pregnant women (n=260) attending antenatal clinics. 10ml of clean- catch midstream urine was collected and cultured on Cysteine Lactose Electrolyte Deficient agar. Bacterial isolates were identified using Gram staining, colony morphology, and biochemical analysis. Antibiotic susceptibility was assessed by Kirby-Bauer disc-diffusion method. Logistic regression was employed for association analysis. *Results:* The prevalence of urinary tract infection was 10.3%. ASB was significantly associated with second trimester (AOR=5.61 [1.40-22.45]; $p<0.05$) and history of urinary tract infection (AOR=12.47 [3.91-39.82]; $p<0.001$). *Escherichia coli* were most prevalent pathogen and gram positive bacteria comprised majority of the overall isolates (63%). The Gram-negatives were more resistant with susceptibility rates of 11.8%, 29.4%, and 35.3% for ampicillin, nalidixic acid and chloramphenicol respectively. The susceptibility rates of Gram-positives were 100%, 80%, 70% for ciprofloxacin and kanamycin, co-trimoxazol and ceftriaxone and gentamycin respectively. Multidrug resistance (≥ 2 drugs) was seen in 81.5% of isolates. *Conclusion:* In generally, pregnant women with ASB may have serious consequences on both mother and fetus. Since in this study the locale specific heterogeneity in ASB prevalence, type and number of bacterial species, dominant bacterial species, antibiotic susceptibility pattern and multidrug resistance was observed. Therefore, it is important to screen all antenatal women for asymptomatic bacteriuria at their first prenatal visit; preferably in first trimester as well as antimicrobial susceptibility testing should be performed for management of ASB among pregnant women.

Keywords: Asymptomatic Bacteriuria, Pregnant Women, Antenatal, Antimicrobial Susceptibility, Ethiopia

1. Introduction

Asymptomatic bacteriuria (ASB) is characterized by the presence of actively multiplying bacteria in the urinary tract excluding the distal urethra without obvious symptoms. Isolation of $\geq 10^5$ cfu/ml of a specific bacterial species in

midstream urine confirms ASB [1-2]. Women are at a higher risk of Urinary tract infection (UTI) and its recurrent episodes [3-4]. It affects up to 10% pregnant women across globe. Local prevalence of ASB is the primary determinant of the incidence of UTI [5]. The apparent reduction in immunity during pregnancy seems to encourage the growth of

opportunistic microorganisms [6-7]. Pregnancy enhances the progression of ASB to symptomatic bacteriuria. This may lead to increased risk of pyelonephritis (20-to-50-fold). These may adversely affect obstetric outcome such as premature delivery, birth weight and fetal mortality rates [4, 8-10]. In spite of this high disease burden, failure to identify ASB during pregnancy is observed among health personnel managing pregnant women [11-13].

The kidneys of the pregnant women excrete waste products of her body as well as those of the foetus. This dual excretory load together with increased incidence of risk factors i.e. ASB associated UTI increase vulnerability to medical complications [14-15]. Approximately 90% of women experience pregnancy associated hydronephrosis. This is progesterone ureteral dilatation begins at week 6 and reaches its peak between weeks 22 to 24. Ureteral dilatation, increased bladder volume and decreased bladder tone, along with decreased ureteral tone, contribute to increased urinary stasis and ureterovesical reflux. Additionally pregnant women develop glycosuria, which may promote bacterial growth in the urine. These factors contribute to the development of asymptomatic UTI during pregnancy [13-15].

The frequency of isolated pathogens, prevalence and antimicrobial resistance patterns varies in different geographical regions [16-20]. Both gram negative and gram-positive bacteria colonize the vaginal introitus and periurethral area. Usually the gram negatives are more prevalent in ASB but the relative prevalence of all other bacterial species causing the ASB varies widely [6, 13, 17-20]. The proper medical interventions including antimicrobial prescription to manage the ASB and associated UTI requires locale specific baseline data related to the type and/strain of the pathogens, their relative prevalence, overall prevalence of ASB and/or UTI and anti-microbial resistance pattern. These information help in providing timely intervention, decrease burden of infection and manage globally increasing trend of anti-microbial resistance. The comprehensive ASB mapping across geographical locales in developing third world countries are indispensable. Therefore this study was conducted to determine the prevalence of ASB and their susceptibility pattern among pregnant women at Mizan Aman town, South-western Ethiopia.

2. Methods

2.1. Study Design and Participants

A cross-sectional study using systematic sampling was performed during September-December of 2017 at General Hospital and Health Center of Mizan Aman town, South-western Ethiopia. Pregnant women ($n=260$; 23.8 ± 3.99 years) attending antenatal clinics (ANC) and without established symptoms of urinary tract disorders/ antibiotic treatment completed the study. Socio-demographic and clinical history data were collected using a structured questionnaire tool. The study was approved by the Research

committee and Publication office, Mizan-Tepi University, Mizan, Ethiopia. Informed written consent was given by the participants.

2.2. Sample Collection and Microbiological Assay

A clean-catch mid-stream urine samples (10 ml) were collected in sterile wide mouthed glass bottles. These were inoculated on Cysteine Lactose Electrolyte Deficient medium (CLED) using calibrated wire loop (0.001 ml) samples. Bacterial counts and (bacterial count $\geq 1 \times 10^5/\text{ml}$) were determined. Single colonies from culture plates with significant bacteriuria were suspended in nutrient broth. These were sub-cultured on Blood agar and MacConkey agar (Oxoid, Ltd) and incubated aerobically at 37°C for 24-48 hrs [21-22].

2.3. Isolation and Identification of Bacterial Pathogens

Bacterial identification was done by their characteristic appearance on medium, Gram staining reaction and pattern of biochemical reactions. Members of the family *Enterobacteriaceae* were identified by indole production, H_2S production, citrate utilization, motility test, urease test, oxidase, carbohydrate utilization tests. For Gram-positive bacteria coagulase and catalase tests were used [21-22].

2.4. Antimicrobial Susceptibility Testing

Antimicrobial susceptibility testing performed on Mueller-Hinton agar plates using Kirby Bauer diffusion technique according to the criteria of the Clinical and Laboratory Standards Institute (CLSI). From a pure culture 3-5 selected colonies of bacteria were inoculated into saline (5 ml) and mixed gently to a homogenous suspension. The turbidity of the suspension was matched to a 0.5 McFarland standard [23-24]. The drugs used in disc diffusion test were in the following concentrations: Ampicillin (AMP) (10 μg), Amoxicillin-Clavulanic Acid (AMC) (30 μg), Ceftriaxone (CRO) (30 μg), Nalidixic Acid (NA) (30 μg), Ciprofloxacin (CIP) (5 μg), Chloramphenicol (C) (30 μg), Cefoxitin (CXT) (30 μg), Doxycycline (DO) (30 μg), Gentamicin (CN) (10 μg), Kanamycin (K) (30 μg) and Trimethoprim-Sulphamethoxazole (TMP- SXT) (1.25/23.75 μg) [21-22].

Culture Media were prepared according to manufacturer's instruction. Sterility of culture media was checked by incubating 5% of the batch at $35 - 37^\circ\text{C}$ overnight and observing the bacterial growth. The quality of culture media and antimicrobial susceptibility were checked using standard reference strain of *P. aeruginosa* (ATCC-27853), *S. aureus* (ATCC-25923) and *E. coli* (ATCC-25922).

2.5. Data Analysis

Statistical analysis was performed by using SPSS version 20.0 (SPSS Inc. Chicago). Descriptive analysis and test of differences were employed. Multiple logistic regression was used to determine the predictors of ASB.

3. Results

3.1. Socio-demographic Characteristics

The socio-demographic characteristics of the participants are summarized in Table 1. Majority of the pregnant women were married (97.7%) and urban dwellers (79.6%). The age

of the pregnant women ranged from 16 to 39 years with median age of 24 years and mean of 23.8 ± 3.99 years. Regarding occupational status of participants, 127 (48.8%) were housewives followed by farmers 48 (18.5%) and government employees 43 (16.6%).

Table 1. Prevalence of asymptomatic UTI and sociodemographic characteristics of pregnant women (N=260) at Mizan-Aman town, South-western Ethiopia, from September-December of 2017.

Characteristics	Number tested (%)	Positive (%)	Negative (%)
Health Facility			
Mizan Aman Hospital	197 (75.8)	24 (12.2)	173 (87.8)
Mizan Aman Health Centre	63 (24.2)	3 (4.8)	60 (95.2)
Ages (years)			
16-20	67 (25.8)	11 (16.4)	56 (83.6)
21-25	120 (46.2)	12 (10.0)	108 (90.0)
26-30	65 (25.0)	4 (6.2)	61 (93.8)
31-35	5 (1.9)	-- (--)	5 (100.0)
36-40	3 (1.2)	-- (--)	3 (100.0)
Marital status			
Married	254 (97.7)	25 (9.8)	229 (90.2)
Other	6 (2.3)	2 (33.4)	4 (66.6)
Address			
Urban	207 (79.6)	22 (10.6)	185 (89.4)
Rural	53 (20.4)	5 (9.4)	48 (90.6)
Occupation			
Farmer	48 (18.5)	6 (12.5)	42 (87.5)
Government employee	43 (16.6)	4 (9.3)	39 (90.7)
Student	8 (3.0)	1 (12.5)	7 (87.5)
Unemployed	12 (4.6)	0 (0)	12 (100)
Housewife	127 (48.8)	15 (11.8)	112 (88.2)
Private business	22 (8.5)	1 (4.5)	21 (95.5)
Monthly income (Birr)			
<500	155 (59.6)	19 (12.2)	136 (87.8)
501-1000	30 (11.5)	3 (10)	27 (90)
1001-1500	23 (8.9)	0 (0)	23 (100)
1501-2000	16 (6.1)	2 (12.5)	14 (87.5)
>2000	36 (13.8)	3 (8.3)	33 (91.7)

3.2. Associated Risk Factors

Prevalence of ASB in this sample of pregnant women was 10.4%. It was associated with second trimester (AOR=5.61 [1.40-22.45]; $p < 0.05$) and history of UTI (AOR=12.47 [3.91-39.82]; $p < 0.001$).

Table 2. Multiple linear regression analysis: Predictors of asymptomatic bacteriuria of pregnant women (N=260) at Mizan-Aman town, South-western Ethiopia, from September-December of 2017.

Variable	Asymptomatic bacteriuria		COR (95%CI)	P-value	AOR (95%CI)	P-value
	No Number (%)	Yes Number (%)				
Age						
16-20	56 (83.6)	11 (16.4)	1			
21-25	108 (90.0)	12 (10.0)	0.57 (0.23-1.36)	0.204		
26-30	61 (93.8)	4 (6.2)	0.33 (0.10-1.11)	0.073		
31-35	5 (100.0)	-- (--)	0.00 (0.00- ---)	0.999		
36-40	3 (100.0)	-- (--)	0.00 (0.00- ---)	0.999		
Trimester						
1	65 (95.6)	3 (4.4)	1		1	
2	85 (81.0)	20 (19.0)	5.10 (1.45-17.90)	0.011	5.61 (1.40-22.45)	0.015
3	83 (95.4)	4 (4.6)	1.04 (0.23-4.83)	0.956	0.80 (0.15-4.18)	0.789
Parity						
Nullipara	43 (93.5)	3 (6.5)	0.35 (0.10-1.26)	0.107	0.36 (0.09-1.53)	0.168
Primary para	105 (93.8)	7 (6.2)	0.33 (0.13-0.84)	0.020	0.47 (0.17-1.30)	0.146
Multipara	85 (83.3)	17 (16.7)	1		1	
History of catheterization						
Yes	7 (63.6)	4 (36.4)	5.61 (1.53-20.63)	0.009	2.57 (0.51-13.06)	0.255

Variable	Asymptomatic bacteriuria		COR (95%CI)	P-value	AOR (95%CI)	P-value
	No Number (%)	Yes Number (%)				
No History of UTI	226 (90.8)	23 (9.2)	1		1	
Yes	13 (56.5)	10 (43.5)	9.95 (3.81-26.02)	0.000	12.47 (3.91-39.82)	0.000
No Marital status	220 (92.8)	17 (7.2)	1		1	
Married	229 (90.2)	25 (9.8)	4.58 (0.80-26.27)	0.088		
Other Residence	4 (66.6)	2 (33.4)	1			
Urban	185 (89.4)	22 (10.6)	1.14 (0.41-3.17)	0.799		
Rural Education	48 (90.6)	5 (9.4)	1			
No formal education	81 (91.0)	8 (9.0)	0.64 (0.18-2.31)	0.497		
Write and read	5 (100)	0 (0)	0.00 (0.00- ---)	0.999		
Elementary (1-8)	49 (80.4)	12 (19.6)	1.59 (0.47-5.43)	0.458		
Secondary (9-12)	72 (96.0)	3 (4.0)	0.27 (0.06-1.29)	0.101		
Higher education (12+)	26 (86.7)	4 (13.3)	1			
Occupation						
Farmer	42 (87.5)	6 (12.5)	1			
Government employee	39 (90.7)	4 (9.3)	0.72 (0.19-2.74)	0.627		
Student	7 (87.5)	1 (12.5)	1.00 (0.10-9.61)	1.000		
Unemployed	12 (100)	0 (0)	0.00 (0.00- ---)	0.999		
Housewife	112 (88.2)	15 (11.8)	0.94 (0.34-2.58)	0.900		
Private business	21 (95.5)	1 (4.5)	0.35 (0.04-3.10)	0.346		

3.3. Isolation and Identification of Bacterial Uropathogens

Gram negative bacteria (63%) was more abundant than the gram positive bacteria (37.0 %) (Table 3). *E. coli* (37.0%) was most prevalent and *Proteus species* (5.88%) least prevalent isolate. *E. coli* (37.0%) and *S. aureus* (25.9%) were predominant gram negative and gram positive isolates respectively (Table 3).

Table 3. : Bacterial isolates and their frequency from urine culture in asymptomatic UTI in Pregnant women at Mizan-Aman town, South-western Ethiopia, from September-December of 2017.

Bacterial isolates	Total (%)
<i>Escherichia coli</i>	10 (37.0)
<i>Klebsiella species</i>	3 (11.2)
<i>Pseudomonas aeruginosa</i>	3 (11.2)
<i>Proteus species</i>	1 (3.70)
<i>Staphylococcus aureus</i>	7 (25.9)
Coagulase negative <i>Staphylococci</i>	3 (11.2)
Total	27 (100)

3.4. Antimicrobial Susceptibility Pattern

Eleven antimicrobial agents were used to determine the susceptibility pattern of the bacterial isolates. The rates of susceptibility of Gram- positives and gram negatives ranged between 10% - 100% and 11.8% - 94.1% respectively. Ciprofloxacin was most effective and ampicillin was the least effective antibiotic against both types of bacterial isolates i.e. gram positive and gram negative (Table 4 and table 5).

Multidrug resistance (resistance to two or more drugs) was identified 81.5% of the bacterial isolates from asymptomatic pregnant women. Gram negative (82.4%) and gram positive (80%) bacteria were multi-drug resistant (Table 6).

Table 4 Antibiotic susceptibility pattern of Gram positive bacterial isolates from urine culture in asymptomatic UTI in pregnant women (N=10) at Mizan-Aman town, South-western Ethiopia, from September-December of 2017.

Species of Bacteria	Number (%) of susceptible strains										
	AMP	CRO	AMC	C	CIP	DO	CN	NA	K	SXT	CXT
<i>S. aureus</i> (n=7)	0 (0)	4 (57.1)	4 (57.1)	4 (57.1)	7 (100)	2 (28.6)	4 (57.1)	2 (28.6)	7 (100)	6 (85.7)	4 (57.1)
<i>CoNS</i> (n=3)	1 (33.4)	3 (100)	1 (33.4)	1 (33.4)	3 (100)	1 (33.4)	3 (100)	1 (33.4)	3 (100)	2 (66.7)	2 (66.7)
Total (n=10)	1 (10)	7 (70)	5 (50)	5 (50)	10 (100)	3 (30)	7 (70)	3 (30)	7 (100)	8 (80)	6 (60)

CoNS=Coagulase negative *Staphylococci*.

AMP: Ampicillin; AMC: Amoxicillin-Clavulanic acid; CRO: Ceftriaxone; C: Chloramphenicol; CIP: Ciprofloxacin; DO: Doxycycline; CN: Gentamicin; K: Kanamycin; SXT: Trimethoprim-sulphamethoxazole; CXT: Cefoxitin and NA: Nalidixic acid.

Table 5. Antibiotic susceptibility pattern of Gram Negative bacterial isolates from urine culture in asymptomatic UTI in pregnant women (N=17) at Mizan-Aman town, South-western Ethiopia, from September-December of 2017.

Species of Bacteria	Number of bacterial strains (%) susceptible to antibiotic tested										
	AMP	AMC	CRO	C	CIP	DO	CN	SXT	NA	K	CXT
<i>E. coli</i> (n=10)	2 (20)	7 (70)	5 (50)	5 (50)	10 (100)	4 (40)	6 (60)	8 (80)	4 (40)	7 (70)	8 (80)
<i>Klebsiella species</i> (n=3)	0 (0)	1 (33.4)	2 (66.7)	1 (33.4)	2 (66.7)	1 (33.4)	0 (0)	2 (66.7)	0 (0)	2 (66.7)	3 (100)
<i>P. aeruginosa</i> (n=3)	0 (0)	3 (100)	0 (0)	0 (0)	3 (100)	1 (33.4)	3 (100)	2 (66.7)	0 (0)	0 (0)	0 (0)
<i>Proteus species</i> (n=1)	0 (0)	1 (100)	1 (100)	0 (0)	1 (100)	1 (100)	1 (100)	1 (100)	1 (100)	1 (100)	1 (100)
Total (n=17)	2 (11.8)	12 (70.6)	8 (47.0)	6 (35.3)	16 (94.1)	7 (41.1)	10 (58.8)	13 (76.5)	5 (29.4)	10 (58.8)	12 (70.6)

AMP: Ampicillin; AMC: Amoxicillin-Clavulanic acid; CRO: Ceftriaxone; C: Chloramphenicol; CIP: Ciprofloxacin; DO: Doxycycline; CN: Gentamicin; K: Kanamycin; SXT: Trimethoprim-sulphamethoxazole; CXT: Cefoxitin and NA: Nalidixic acid.

Table 6. Multidrug resistance pattern of bacterial isolates from urine culture in asymptomatic UTI in pregnant women (N=27) at Mizan-Aman town, South-western Ethiopia, from September-December of 2017.

Antimicrobial pattern							
Bacterial isolate	Total (%)	R0	R1	R2	R3	R4	≥R5
Gram negative	17 (63.0)	0 (0)	3 (17.6)	0 (0)	1 (5.9)	3 (17.6)	10 (58.8)
<i>Escherichia coli</i>	10 (58.8)	0 (0)	2 (20)	0 (0)	0 (0)	3 (30)	5 (50)
<i>Klebsiella species</i>	3 (17.6)	0 (0)	0 (0)	0 (0)	1 (33.3)	0 (0)	2 (66.7)
<i>Pseudomonas aeruginosa</i>	3 (17.6)	0 (0)	0 (0)	0 (0)	0 (0)	0 (0)	3 (100)
<i>Proteus species</i>	1 (5.9)	0 (0)	1 (100)	0 (0)	0 (0)	0 (0)	0 (0)
Gram Positive	10 (37.0)	0 (0)	2 (20)	1 (10)	0 (0)	1 (10)	6 (60)
<i>Staphylococcus aureus</i>	7 (70)	0 (0)	1 (14.3)	1 (14.3)	0 (0)	1 (14.3)	4 (57.1)
Coagulase negative staphylococci	3 (30)	0 (0)	1 (33.3)	0 (0)	0 (0)	0 (0)	2 (66.7)
Total	27 (100)	0 (0)	5 (18.5)	1 (3.7)	1 (3.7)	4 (14.8)	16 (59.3)

R0- No antibiotic resistance, R1- Resistance to one, R2-Resistance to two, R3-Resistance to three, R4- Resistance to four, ≥R5- resistance to five and more drugs.

4. Discussions

The findings reinstated the working hypothesis about the locale specific prevalence rate of ASB, types of bacteria causing ASB and their differential susceptibility pattern among pregnant women. Globally, the prevalence rate of 3.6%-37.1% has been reported [23-24]. The prevalence of ASB among pregnant women in this study was within the national range (7%-18.8%) reported in Ethiopian women of other regions [19, 25-27]. This difference may be related to geographical location, environmental conditions, social and personal hygiene and/or negligence of ASB by the health care system. This preliminary prevalence in double digit indicates neglect of ASB related health issues in this South Ethiopian region. The course correction measures may be implemented. The numbers of bacterial species involved in ASB have a geographical outline. As against our finding of six bacterial isolates, previous literatures on Ethiopian populations enlist 5-9 bacterial species causing ASB [17, 28-29]. The literature is not unanimous on the most prevalent bacteria even in Ethiopian population. Though majority of the studies including our work found *E. coli* to be the most prevalent in causing ASB [12, 17, 28-31]. The physio-anatomical changes and difficulty in personal hygiene maintenance during pregnancy may increase the risk of urogenital colonization by *E. coli*. Moreover increased level of amino acids and lactose during pregnancy encourage growth of *E. coli* [30-31]. *Coagulase negative Staphylococcus* (32.6%) was found to be the leading cause

of ASB in Hawassa region [26]. Most of the previous studies failed to identify infection by *Proteus species* in Ethiopians pregnant women [19, 28, 31-32]. This regional pattern with respect to the types, number, and most prevalent bacteria causing ASB highlight the challenge in managing it.

Similar to previous reports, place of residence, gravidity, occupation, and marital status was not found to be associated with ASB (Table 2) [31, 33]. However, there are contradictory evidences regarding relationship between maternal age and ASB [29-32, 34-35]. Some studies reported association with increasing age [34], while others found it to be related with younger age group [30-31]. This future studies should try to look into the aspect. In our study, contrary to previous studies in Ethiopia [19, 26, 31], trimester (gestational age) was associated with prevalence of ASB. Pregnant women in second trimester had high likelihood of testing positive for significant bacteriuria ($p<0.05$) compared to pregnant women in the first trimester. This susceptibility during this period may be related to ureteral dilatation. The dilatation stimulates production of progesterone and estrogen which lowers the tone of the uterus [12, 36]. Similar to previous findings, the prevalence of ASB in pregnant women was associated with history of UTI ($p<0.001$) [19, 31]. The presence of resistant strains may be implicated in this relationship.

Antimicrobial resistance is an area of increasing concern in the management of UTI [31-32]. The prescription of antibiotics without proper test of susceptibility is a contributing factor. The drugs with higher susceptibility

should be used for management of ASB. Therefore, Ciprofloxacin, kanamycin and trimethoprim-sulphamethoxazole may be prescribed in this South western Ethiopian pregnant women population to manage ASB caused by gram positive bacteria (Table 4). Ciprofloxacin, trimethoprim- sulphamethoxazole, amoxicillin-clavulnic acid and cefoxitin may be used to manage ASB caused by gram negative bacteria, because of the higher susceptibility bacteria to these antibiotics. The higher resistance for ampicillin (88.2%), nalidixic acid (70.6%) and chloramphenicol (64.7%) is similar to previous reports [15]. Earlier exposure of the isolates to these drugs may be related to increased resistance development. Similar to previous reports, multi drug resistance (MDR= resistance to ≥ 2 drugs) was observed in the overwhelming majority (81.5%) of the bacterial isolate (Table 6) [19, 31-32]. The prescription of antibiotics without susceptibility pattern analysis and unregulated over-the-counter sale of antibiotics are most likely reason for increasing resistance.

5. Conclusion

In generally, pregnant women with ASB may have serious consequences on both mother and fetus. Since in this study the locale specific heterogeneity in ASB prevalence, type and number of bacterial species, dominant bacterial species, antibiotic susceptibility pattern and multidrug resistance was observed. Proper clinical management of ASB depends on locale specific information like prevalence; type and number of bacterial species, dominant bacterial species, antibiotic susceptibility pattern and multidrug resistance. Therefore, it is important to screen all antenatal women for asymptomatic bacteriuria at their first prenatal visit; preferably in first trimester as well as antimicrobial susceptibility testing should be performed for management of ASB among pregnant women.

Data Availability

All relevant data are within the article, but any additional data required are available from the corresponding author upon request.

Ethical Approval

Ethical approval and clearance letter of permission were obtained from ethical clearance committee of College of Health Science and Medicine, Mizan-Tepi University. An official letter written from Mizan-Tepi University Teaching Hospital Chief Executive Office was distributed to each study ward and area.

Conflict of Interest

All the authors do not have any possible conflicts of interest.

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Authors' Contributions

AK conceived, designed, performed data collection, data analysis, drafted the manuscript. MT interpreted the results, reviewed and approved the manuscript. AK drafted the manuscript.

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